



LipidLaunch™ BODIPY SM-102 LNP Kit (Loadable)

Item No. 702760

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GENERAL INFORMATION

Materials Supplied

Kit will arrive packaged as a -80°C kit. After opening the kit, store individual components as stated below.

Item Number	Item Name	Quantity/Size	Storage Temperature
400774	LipidLaunch™ BODIPY SM-102 LNP (Loadable)	2 vials	-80°C
400649	LNP Dilution Buffer A (1X)	1 vial/20 ml	4°C
400812	LNP Encapsulation Buffer Tablet	1 tablet	RT

If any of the items listed above are damaged or missing, please contact our Customer Service department at (800) 364-9897 or (734) 971-3335. We cannot accept any returns without prior authorization.



WARNING: THIS PRODUCT IS FOR RESEARCH ONLY - NOT FOR HUMAN OR VETERINARY DIAGNOSTIC OR THERAPEUTIC USE.

Safety Data

This material should be considered hazardous until further information becomes available. Do not ingest, inhale, get in eyes, on skin, or on clothing. Wash thoroughly after handling. Before use, the user must review the complete Safety Data Sheet, which has been sent *via* email to your institution.

Precautions

Please read these instructions carefully before beginning this assay.

The reagents in this kit have been tested and formulated to work exclusively with Cayman Chemical's LipidLaunch™ BODIPY SM-102 LNP Kit (Loadable).

If You Have Problems

Technical Service Contact Information

Phone: 888-526-5351 (USA and Canada only) or 734-975-3888

Email: techserv@caymanchem.com

In order for our staff to assist you quickly and efficiently, please be ready to supply the lot number of the kit (found on the outside of the box).

Storage and Stability

Kit components should be stored as directed in the Materials Supplied section (see page 3) and used before the expiration date indicated on the outside of the box.

Materials Needed But Not Supplied

1. A source of nuclease-free water is recommended. Pure water - glass-distilled or deionized - may not be acceptable.
2. Adjustable pipettes; multichannel or repeating pipettor recommended

INTRODUCTION

Background

Lipid nanoparticles (LNPs) are a subset of lipid-based drug delivery (LBDD) systems that utilize ionizable cationic lipids, such as SM-102, for the delivery of nucleic acid payloads to cells.^{1,2} LNPs are typically composed of four types of lipids: a cationic or ionizable cationic lipid, a helper phospholipid, a PEGylated lipid, and cholesterol. Release of LNP cargo into target cells is heavily influenced by the ionizable cationic lipid component, which undergoes protonation in the acidic environment of the endosomes, resulting in membrane disruption and release of cargo into the cell.³ SM-102 is an ionizable cationic aminolipid that has been used in the generation of LNPs for the delivery of cargos, such as mRNA and plasmid DNA *in vitro* and *in vivo*.^{4,5} In mice, SM-102-containing LNPs have been shown to accumulate in the lungs and spleen.⁶ Cayman's LipidLaunch™ BODIPY SM-102 LNP Kit (Loadable) uses the ionizable cationic aminolipid SM-102 to facilitate efficient payload delivery in research models with reduced toxicity compared to traditional methods.

About This Kit

Cayman's LipidLaunch™ BODIPY SM-102 LNP Kit (Loadable) provides cargo-ready, empty, BODIPY-labelled SM-102 LNPs. Following resuspension, these LNPs are capable of rapid encapsulation of nucleic acids, without the need for incubation steps, enabling subsequent delivery to target cells or other downstream research applications. LipidLaunch™ LNPs (Loadable) demonstrate very low toxicity compared to traditional lipid-based transfection methods.

Conventionally-loaded LNPs typically require specialized equipment to ensure consistent control of particle size, while also requiring large reaction volumes. LipidLaunch™ BODIPY SM-102 LNPs (Loadable) enable users to encapsulate cargo in low reaction volumes offering greater flexibility in the scale of preparation while preserving costly nucleic acid cargo.

LipidLaunch™ BODIPY SM-102 LNP (Loadable) performance is dependent on a variety of factors, including cargo type and cell line, so optimization of these parameters is necessary. To aid in optimization, consider using Cayman's LipidLaunch™ LNP Screening Set (Loadable) (Item No. 502868) or LipidLaunch™ SM-102 LNPs preloaded with luciferase, mCherry, or GFP mRNA (Item Nos. 39318 | 39319 | 39320).

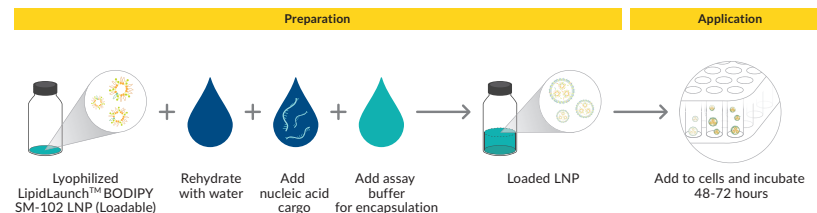


Figure 1. Protocol summary

Reagent Preparation

1. LipidLaunch™ BODIPY SM-102 LNP (Loadable) - (Item No. 400774)

Reconstitute one vial with 200 µl nuclease-free water. Gently swirl the vial 2-3 times to fully resuspend. Each vial of reconstituted BODIPY SM-102 LNPs provides a sufficient volume to transfect up to 50 wells using 24-well plates. Reconstituted LNPs are stable for up to one week when stored at 4°C. Do not freeze.

2. LNP Dilution Buffer A (1X) - (Item No. 400469)

This vial contains 20 ml LNP Dilution Buffer A (1X). This optional reagent is intended for optimizing the cargo:LNP loading ratio prior to encapsulation (see Cargo Encapsulation, page 10).

3. Encapsulation Buffer Preparation

Dissolve the LNP Encapsulation Buffer Tablet (Item No. 400812) in 10 ml of sterile-filtered nuclease-free water. The Encapsulation Buffer will be stable for six months when stored at 4°C.

Reagent Protocol

General Information

- LNP loading can be performed at room temperature.
- If not using loaded LNPs immediately, store at 4°C. Do not freeze.
- It is recommended to use loaded LNPs within one week. Stability of loaded LNPs may vary depending on the cargo type and handling prior to loading.
- Loaded LNPs can be diluted directly into culture medium or a neutral buffer of choice.
- If transfecting cells, optimal expression is typically observed using 1-3 µl of loaded LNPs per 100 µl of medium.
- Loaded LNPs may be sterile-filtered using 0.2 µm polyethersulfone (PES) syringe filters without affecting performance.

1. Cargo Encapsulation

- Prepare cargo in nuclease-free water. If using RNA, it is recommended to optimize by initially loading with 10-100 ng/μl.
- Loadable LNPs may be optionally diluted with LNP Dilution Buffer A (1X) prior to encapsulation.
- LNPs are loaded by adding reagents in a specific order:
 1. Gently mix loadable LNPs with cargo.
 2. Add Encapsulation Buffer to complete loading.
- Cargo and LNP volumes can be varied, however, **it is necessary that the volume of Encapsulation Buffer added is one-third the combined volume of cargo and LNP.**
- Table 1, below, shows an example of recommended loading volumes. Scale accordingly.

Procedure	Volume
Add Loadable LNP	20 μl
Add Cargo	4 μl
Pipette up and down to mix	
Add Encapsulation Buffer	8 μl
Pipette up and down to mix	

Table 1. Recommended initial loading conditions

2. Characterization

A variety of techniques are available to characterize LNPs prior to *in vitro* or *in vivo* use.

Attribute	Assay(s)
Particle size and distribution	Dynamic light scattering (DLS)
Zeta potential	Laser doppler electrophoresis
Lipid quantification and integrity	RP-HPLC, SE-HPLC, IP-HPLC
Encapsulation efficiency	Fluorescent dyes (RiboGreen); UV spectroscopy with Triton X-100
LNP morphology	Microscopy (cryo TEM, ESEM, AFM)
Translation or knockdown analyses	Cell-based reporter assays, Western blotting

Table 2. LNP attributes and corresponding assays. Adapted from Schoenmaker, L., *et al.*⁷

Example Data

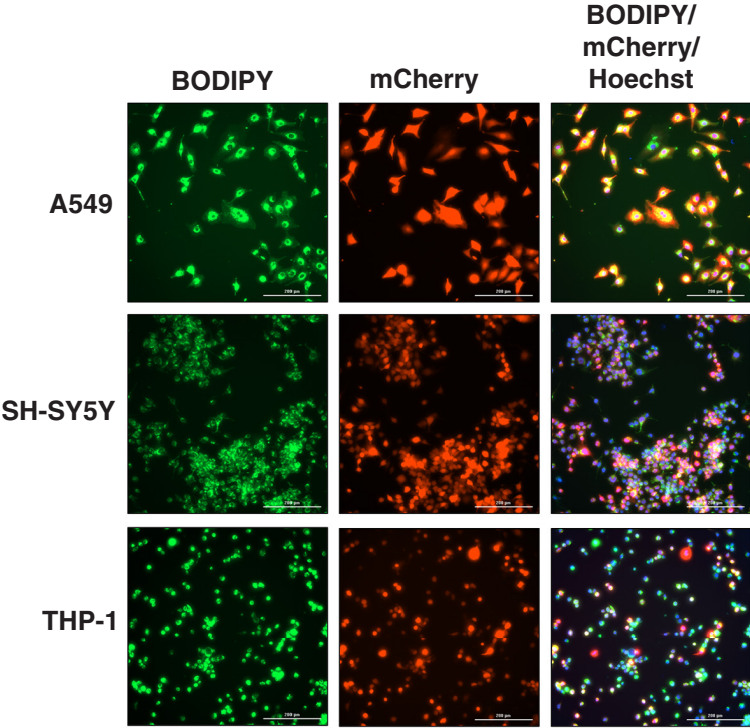


Figure 2. Typical transfection results with mCherry mRNA-loaded LipidLaunch™ BODIPY SM-102 LNPs (Loadable). A549, SH-SY5Y, and THP-1 cells were transfected with loadable LNPs following encapsulation of mCherry mRNA. Fluorescence images were collected at 20X magnification 72 hours later (BODIPY (left), mCherry (center), BODIPY/mCherry/Hoechst (right); Bar size: 200 µm).

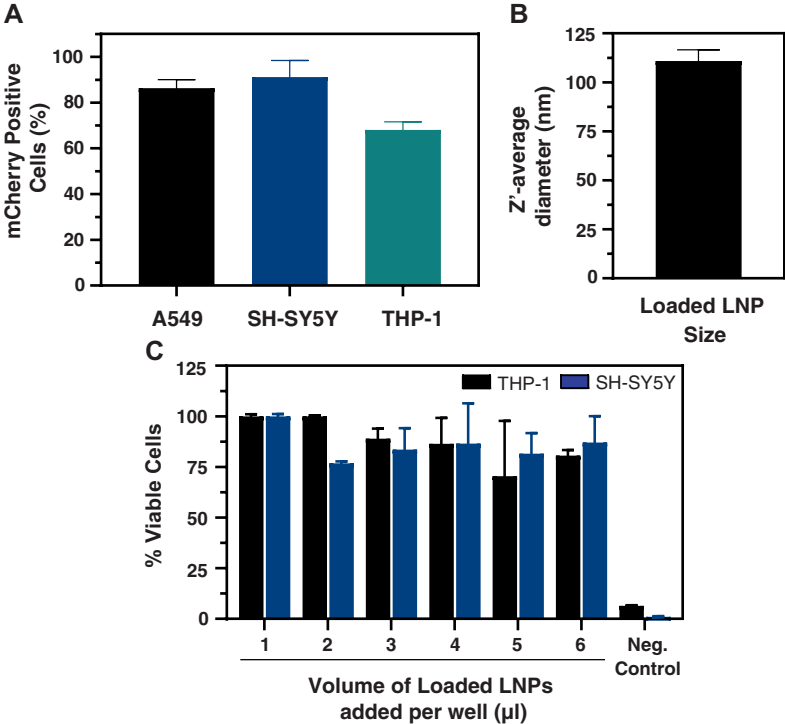


Figure 3. Characterization of mCherry mRNA-loaded LipidLaunch™ BODIPY SM-102 LNPs (Loadable) and their impact on cell viability. (A) The transfection efficiency of the experiment shown in Fig. 2 was determined by scoring the percentage of mCherry-positive cells. (B) The average loaded particle size was determined via DLS. (C). Cell viability was determined via the Resazurin Cell Viability Assay Kit (Item No. 702540) 72 hours following transfection with mCherry mRNA-loaded LNPs without changing the medium (50 ng mRNA, 100 µl medium per well).

Troubleshooting

Problem	Possible Causes	Recommended Solutions
Poor encapsulation efficiency	A. RNA/cargo is degraded B. Reagents added in incorrect order C. RNA was prepared incorrectly D. Suboptimal cargo:LNP ratio	A. Use fresh RNA/cargo B. Ensure cargo and LNP are mixed prior to adding the Encapsulation Buffer C. Dissolve RNA in nuclease-free water D. Optimize cargo concentration and cargo:LNP ratio

References

- Mitchell, M.J., Billingsley, M.M., Haley, R.M., *et al.* *Nat. Rev. Drug Discov.* **20(2)**, 101-124 (2021).
- Viegas, C., Patrício, A.B., Prata, J.M., *et al.* *Pharmaceutics* **15**, 1593 (2023).
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Warranty and Limitation of Remedy

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