Maresin 1 ELISA Kit

Item No. 501150

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GENERAL INFORMATION

Materials Supplied

Item Number	Item	96 wells Quantity/Size
401152	Maresin 1 ELISA Antiserum	1 vial/100 dtn
401150	Maresin 1 AChE Tracer	1 vial/100 dtn
401154	Maresin 1 ELISA Standard	1 vial
400060	ELISA Buffer Concentrate (10X)	2 vials/10 ml
400062	Wash Buffer Concentrate (400X)	1 vial/5 ml
400035	Polysorbate 20	1 vial/3 ml
400004/400006	Mouse Anti-Rabbit IgG Coated Plate	1 plate
400012	96-Well Cover Sheet	1 cover
400050	Ellman's Reagent	3 vials/100 dtn
400040	ELISA Tracer Dye	1 vial
400042	ELISA Antiserum Dye	1 vial

If any of the items listed above are damaged or missing, please contact our Customer Service department at (800) 364-9897 or (734) 971-3335. We cannot accept any returns without prior authorization.

WARNING: THIS PRODUCT IS FOR RESEARCH ONLY - NOT FOR HUMAN OR VETERINARY DIAGNOSTIC OR THERAPEUTIC USE.

Safety Data

This material should be considered hazardous until further information becomes available. Do not ingest, inhale, get in eyes, on skin, or on clothing. Wash thoroughly after handling. Before use, the user must review the complete Safety Data Sheet, which has been sent *via* email to your institution.

Precautions

Please read these instructions carefully before beginning this assay.

The reagents in this kit have been tested and formulated to work exclusively with Cayman Chemical's Maresin 1 ELISA Kit. This kit may not perform as described if any reagent or procedure is replaced or modified.

When compared to quantification by LC-MS or GC-MS, it is not uncommon for immunoassays to report higher analyte concentrations. While LC-MS or GC-MS analyses typically measure only a single compound, antibodies used in immunoassays sometimes recognize not only the target molecule, but also structurally related molecules, including biologically relevant metabolites. In many cases, measurement of both the parent molecule and metabolites is more representative of the overall biological response than is the measurement of a short-lived parent molecule. It is the responsibility of the researcher to understand the limits of both assay systems and to interpret their data accordingly.

Researchers are advised to validate results from this ELISA with a secondary detection method or to clearly report the limitations of this ELISA in their publications.

If You Have Problems

Technical Service Contact Information

Phone: 888-526-5351 (USA and Canada only) or 734-975-3888

Email: techserv@caymanchem.com

In order for our staff to assist you quickly and efficiently, please be ready to supply the lot number of the kit (found on the outside of the box).

Storage and Stability

This kit will perform as specified if stored as directed at -80°C and used before the expiration date indicated on the outside of the box.

Materials Needed But Not Supplied

1. A plate reader capable of measuring absorbance between 405-420 nm.
2. Adjustable pipettes and a repeating pipettor.
3. A source of ultrapure water, with a resistivity of 18.2 MΩ·cm and total organic carbon (TOC) levels of <10 ppb, is recommended. Pure water - glass-distilled or deionized - may not be acceptable. *NOTE: UltraPure Water is available for purchase from Cayman (Item No. 400000).*
4. Materials used for **Sample Preparation** (see page 10).

INTRODUCTION

Background

Maresins (macrophage mediators in resolving inflammation) comprise part of a class of potent anti-inflammatory and pro-resolving mediators. These mediators are synthesized by macrophages in the presence of docosahexaenoic acid (DHA). Maresin 1 (MaR1) is produced *via* oxidation of endogenous DHA by 12-lipoxygenase. The resulting 14-hydroperoxy docosahexaenoic acid subsequently produces a 13(S),14(S)-epoxide intermediate, which is then hydrolyzed to MaR1.[1] MaR1 has been shown to reduce polymorphonuclear neutrophil (PMN) infiltration in murine peritonitis, enhance efferocytosis, inhibit TRPV1 currents in neurons, and reduce both inflammation- and chemotherapy-induced neuropathic pain in mice.[2] Additionally, systemic treatment with MaR1 attenuated both DSS- and TNBS-induced colitis in the mouse model. This resulted in a significant decrease in inflammatory mediator synthesis, colonic cell infiltration, and reactive oxygen species production.[3] Although circulating levels of MaR1 may be quite low, MaR1 can be detected in urine.[4]

About This Assay

Cayman's MaR1 ELISA Kit is a competitive assay that can be used for quantification of MaR1. This assay has been tested in urine. The ELISA typically displays an 50% B/B_0 value of approximately 42.8 pg/ml and a detection limit (80% B/B_0) of approximately 9.6 pg/ml.

Description of AChE Competitive ELISAs

This assay is based on the competition between free MaR1 and an MaR1 Tracer (MaR1 linked to acetylcholinesterase (AChE)) for a limited number of MaR1-specific rabbit antiserum binding sites. The concentration of the MaR1 Tracer is held constant while the concentration of free MaR1 (standard or sample) varies. Thus, the amount of MaR1 Tracer that is able to bind to the rabbit antiserum will be inversely proportional to the concentration of free MaR1 in the well. This rabbit antiserum-MaR1 (either free or tracer) complex binds to the mouse monoclonal anti-rabbit IgG that has been previously attached to the well. The plate is washed to remove any unbound reagents and then Ellman's Reagent (which contains the substrate to AChE) is added to the well. The product of this enzymatic reaction has a distinct yellow color and absorbs strongly at 414 nm. The intensity of this color, determined spectrophotometrically, is proportional to the amount of MaR1 Tracer bound to the well, which is inversely proportional to the amount of free MaR1 present in the well during the incubation; or

$$\text{Absorbance} \propto \text{[Bound MaR1 Tracer]} \propto 1/\text{[MaR1]}$$

A schematic of this process is shown below in Figure 1.

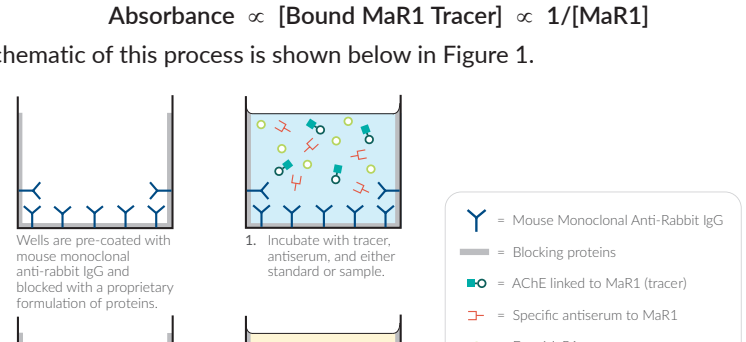


Figure 1. Schematic of the AChE ELISA

Definition of Key Terms

Blank: background absorbance caused by Ellman's Reagent. The blank absorbance should be subtracted from the absorbance readings of all the other wells, including NSB wells.

Total Activity: total enzymatic activity of the AChE-linked tracer. This is analogous to the specific activity of a radioactive tracer.

NSB (Non-Specific Binding): non-immunological binding of the tracer to the well. Even in the absence of specific antibody a very small amount of tracer still binds to the well; the NSB is a measure of this low binding. Do not forget to subtract the Blank absorbance values.

B_0 (Maximum Binding): maximum amount of the tracer that the antibody can bind in the absence of free analyte.

%B/B_0 (%Bound/Maximum Bound): ratio of the absorbance of a particular sample or standard well to that of the maximum binding (B_0) well.

Dynamic Range: the range in which the analyte is reliably quantifiable.

Dtn: determination, where one dtn is the amount of reagent used per well.

Cross Reactivity: numerical representation of the relative reactivity of this assay towards structurally related molecules as compared to the primary analyte of interest. Biomolecules that possess similar epitopes to the analyte can compete with the assay tracer for binding to the primary antibody. Substances that are superior to the analyte in displacing the tracer result in a cross reactivity that is greater than 100%. Substances that are inferior to the primary analyte in displacing the tracer result in a cross reactivity that is less than 100%. Cross reactivity is calculated by comparing the mid-point (50% B/B_0) value of the tested molecule to the mid-point (50% B/B_0) value of the primary analyte when each is measured in assay buffer using the following formula:

$$\text{\% Cross Reactivity} = \left[\frac{\text{50\% B/B}_0 \text{ value for the primary analyte}}{\text{50\% B/B}_0 \text{ value for the potential cross reactant}} \right] \text{x } 100\%$$

PRE-ASSAY PREPARATION

NOTE: Water used to prepare all ELISA reagents and buffers must be deionized and free of trace organic contaminants ('UltraPure'). Use activated carbon filter cartridges or other organic scavengers. Glass distilled water (even if double distilled), HPLC-grade water, and sterile water (for injections) are not adequate for ELISA. UltraPure water may be purchased from Cayman (Item No. 400000).

Buffer Preparation

Store all diluted buffers at 4°C; they should be stable for about two months.

1. **ELISA Buffer Preparation**

   Dilute the contents of one vial of ELISA Buffer Concentrate (10X) (Item No. 400060) with 90 ml of UltraPure water. Be certain to rinse the vial to remove any salts that may have precipitated. *NOTE: It is normal for the concentrated buffer to contain crystalline salts after thawing. These will completely dissolve upon dilution with water.*

2. **Wash Buffer Preparation**

   5 ml vial Wash Buffer Concentrate (400X) (Item No. 400062): Dilute to a total volume of 2 liters with UltraPure water and add 1 ml of Polysorbate 20 (Item No. 400035).

Smaller volumes of Wash Buffer can be prepared by diluting the Wash Buffer Concentrate 1:400 and adding Polysorbate 20 (0.5 ml/liter of Wash Buffer).

NOTE: Polysorbate 20 is a viscous liquid and cannot be measured by a regular pipette. A positive displacement pipette or a syringe should be used to deliver small quantities accurately.

Sample Preparation

Testing for Interference

This assay has been tested using human urine. Other sample types should be checked for interference to evaluate the need for sample purification before embarking on a large number of sample measurements. To test for interference, dilute one or two test samples to obtain at least two different dilutions of each sample between approximately 186.1 and 9.6 pg/ml (*i.e.*, between 20-80% B/B_0, which is the linear portion of the standard curve). If the two different dilutions of the sample show good correlation (differ by 20% or less) in the final calculated MaR1 concentration, extraction is not required. If you do not see good correlation of the different dilutions, extraction is advised. If you choose to extract your samples, references 4, 6, and 7 may provide useful guidance.

General Precautions

- All samples must be free of organic solvents prior to assay.
- Samples should be assayed immediately after collection; samples that cannot be assayed immediately should be stored at -80°C. Degradation of samples has been noticed over one week time period in urine samples stored at -80°C.
- Samples of rabbit origin may contain antibodies which interfere with the assay by binding to the mouse anti-rabbit plate. We recommend that all rabbit samples be purified prior to use in this assay.
- AEBSF (Pefabloc SC®) and PMSF inhibit AChE. Samples containing these protease inhibitors should not be used in this assay.

Urine

MaR1 can be measured in urine without extraction. Alternatively, urine samples may be treated with β-glucuronidase prior to assay, as described by Skarke, C. *et al.*[4] Treatment of urine with β-glucuronidase will remove of a sugar residue from MaR1, improving the recognition of urinary MaR1 by the MaR1 Antiserum.

If you wish to treat urine with β-glucuronidase prior to immunoassay, the following protocol is recommended.

1. Dilute 1 ml of urine with 1 ml of 1 M sodium acetate, pH 5.0.
2. Add 100 µl of β-glucuronidase from *Helix pomatia*.
3. Incubate overnight at 37°C.
4. Neutralize by the addition of 1 ml of 1 M potassium phosphate, pH 7.4. These neutralized samples can be used directly in the ELISA.

Sample Matrix Properties

Linearity

A human urine sample with a biologically relevant level of MaR1 was checked at multiple dilutions using the MaR1 ELISA. The results are shown in the table below.

Dilution	Concentration (pg/ml)	Dilution Linearity (%)
Neat	186.1	89
1:2	208.8	100
1:4	211.0	101
1:6	209.7	100
1:8	204.2	98
1:12	212.5	102

Table 1. Dilutional Linearity of human urine in the MaR1 ELISA

Spike and Recovery

Human urine was spiked with MaR1, diluted as described in the **Sample Preparation** section and analyzed using the MaR1 ELISA Kit. The results are shown below. The y-intercept corresponds to the amount of endogenous MaR1 in the urine sample. Error bars represent standard deviations obtained from multiple dilutions of each sample.

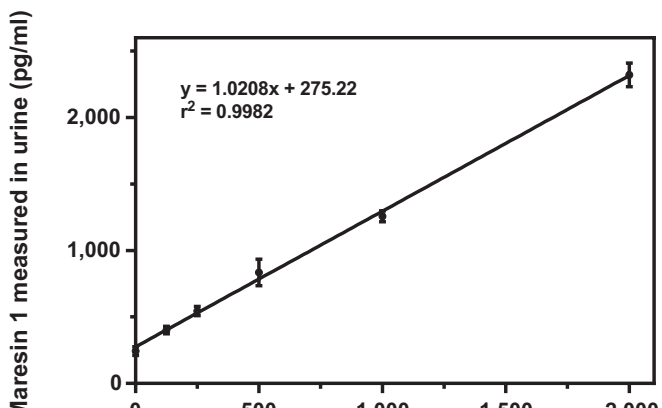


Figure 2. Spike and recovery in the MaR1 ELISA

Parallelism

To assess parallelism, a human urine sample was checked at multiple dilutions by the MaR1 ELISA Kit. Concentrations were plotted as a function of the sample dilution. The results are shown below. Parallelism of the curves demonstrates that the antigen binding characteristics are similar enough to allow the accurate determination of native analyte levels in diluted human urine sample.

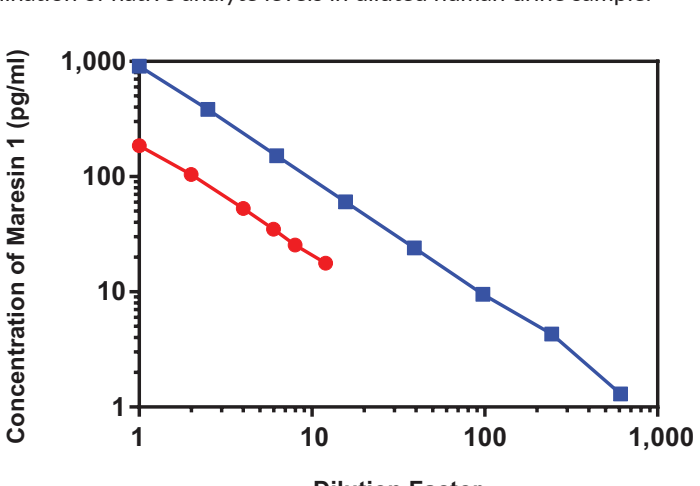


Figure 3. Parallelism of sample matrices in the MaR1 ELISA

ASSAY PROTOCOL

Preparation of Assay-Specific Reagents

Maresin 1 ELISA Standard

Equilibrate a pipette tip in ethanol by repeatedly filling and expelling the tip with ethanol several times. Using the equilibrated pipette tip, transfer 100 µl of the MaR1 ELISA Standard (Item No. 401154) into a clean test tube, then dilute with 900 µl of UltraPure water. The concentation of this solution (the bulk standard) will be 10 ng/ml.

NOTE: If assaying culture medium samples that have not been diluted with ELISA Buffer, culture medium should be used in place of ELISA Buffer for dilution of the standard curve.

To prepare the standard for use in ELISA: Obtain eight clean test tubes and number them #1 through #8. Aliquot 900 µl ELISA Buffer to tube #1 and 600 µl ELISA Buffer to tubes #2-8. Transfer 100 µl of the bulk standard (10 ng/ml) to tube #1 and mix thoroughly. Serially dilute the standard by removing 400 µl from tube #1 and placing in tube #2; mix thoroughly. Next, remove 400 µl from tube #2 and place it into tube #3; mix thoroughly. Repeat this process for tubes #4-8. The diluted standards may be stored at 4°C for no more than 24 hours.

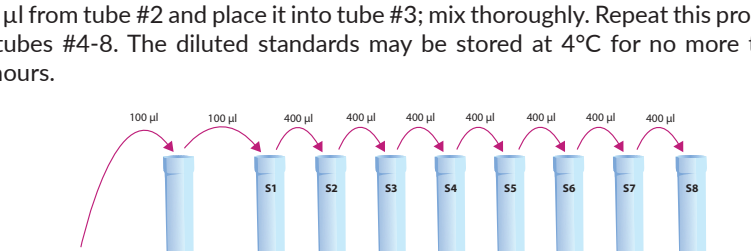


Figure 4. Preparation of the MaR1 standards

Maresin 1 AChE Tracer

Reconstitute the MaR1 AChE Tracer as follows:

100 dtn MaR1 AChE Tracer (Item No. 401150): Reconstitute with 6 ml ELISA Buffer.

Store the reconstituted MaR1 AChE Tracer at 4°C *(do not freeze!)* and use within four weeks. A 20% surplus of tracer has been included to account for any incidental losses.

Tracer Dye Instructions (optional)

This dye may be added to the tracer, if desired, to aid in visualization of tracer-containing wells. Add the dye to the reconstituted tracer at a final dilution of 1:100 (add 60 µl of dye to 6 ml tracer).

Maresin 1 ELISA Antiserum

Reconstitute the MaR1 ELISA Antiserum as follows:

100 dtn MaR1 ELISA Antiserum (Item No. 401152): Reconstitute with 6 ml ELISA Buffer.

Store the reconstituted MaR1 ELISA Antiserum at 4°C. It should be stable for at least four weeks. A 20% surplus of antiserum has been included to account for any incidental losses.

Antiserum Dye Instructions (optional)

This dye may be added to the antibody, if desired, to aid in visualization of antibody-containing wells. Add the dye to the reconstituted antibody at a final dilution of 1:100 (add 60 µl of dye to 6 ml antibody).

Plate Set Up

The 96-well plate(s) included with this kit is supplied ready to use. It is not necessary to rinse the plate(s) prior to adding the reagents. *NOTE: If you do not need to use all the strips at once, place the unused strips back in the plate packet and store at 4°C. Be sure the packet is sealed with the desiccant inside.*

Each plate or set of strips must contain a minimum of two blanks (Blk), two non-specific binding wells (NSB), two maximum binding wells (B_0), and an eight point standard curve run in duplicate. *NOTE: Each assay must contain this minimum configuration in order to ensure accurate and reproducible results.* Each sample should be assayed at two dilutions and each dilution should be assayed in duplicate. For statistical purposes, we recommend assaying samples in triplicate.

A suggested plate format is shown below in Figure 5. The user may vary the location and type of wells present as necessary for each particular experiment. The plate format provided below has been designed to allow for easy data analysis using a convenient spreadsheet offered by Cayman (see page 21, for more details). We suggest you record the contents of each well on the template sheet provided (see page 29).

	1	2	3	4	5	6	7	8	9	10	11	12
A	Blk	S1	S1	1	1	1	9	9	9	17	17	17
B	Blk	S2	S2	2	2	2	10	10	10	18	18	18
C	NSB	S3	S3	3	3	3	11	11	11	19	19	19
D	NSB	S4	S4	4	4	4	12	12	12	20	20	20
E	B_0	S5	S5	5	5	5	13	13	13	21	21	21
F	B_0	S6	S6	6	6	6	14	14	14	22	22	22
G	B_0	S7	S7	7	7	7	15	15	15	23	23	23
H	TA	S8	S8	8	8	8	16	16	16	24	24	24

Blk - Blank
TA - Total Activity
NSB - Non-Specific Binding
B_0 - Maximum Binding
S1-S8 - Standards 1-8
1-24 - Samples

Figure 5. Sample plate format

Performing the Assay

Pipetting Hints

- Use different tips to pipette each reagent.
- Before pipetting each reagent, equilibrate the pipette tip in that reagent (*i.e.*, slowly fill the tip and gently expel the contents, repeat several times).
- Do not expose the pipette tip to the reagent(s) already in the well.

Addition of the Reagents

1. **ELISA Buffer**

   Add 100 µl ELISA Buffer to NSB wells. Add 50 µl ELISA Buffer to B_0 wells. If culture medium was used to dilute the standard curve, substitute 50 µl of culture medium for ELISA Buffer in the NSB and B_0 wells (*i.e.*, add 50 µl culture medium to NSB and B_0 wells and 50 µl ELISA Buffer to NSB wells).

2. **Maresin 1 ELISA Standard**

   Add 50 µl from tube #8 to both of the lowest standard wells (S8). Add 50 µl from tube #7 to each of the next two standard wells (S7). Continue with this procedure until all the standards are aliquoted. The same pipette tip should be used to aliquot all the standards. Before pipetting each standard, be sure to equilibrate the pipette tip in that standard.

3. **Samples**

   Add 50 µl of sample per well. Each sample should be assayed at a minimum of two dilutions. Each dilution should be assayed in duplicate (triplicate recommended).

4. **Maresin 1 AChE Tracer**

   Add 50 µl to each well *except* the TA and the Blk wells.

5. **Maresin 1 ELISA Antiserum**

   Add 50 µl to each well *except* the TA, the NSB, and the Blk wells.

Incubation of the Plate

Cover each plate with plastic film (Item No. 400012) and incubate for 18 hours at 4°C.

Development of the Plate

1. Reconstitute Ellman's Reagent immediately before use (20 ml of reagent is sufficient to develop 100 wells):

   100 dtn vial Ellman's Reagent (Item No. 400050): Reconstitute with 20 ml of UltraPure water.

NOTE: Reconstituted Ellman's Reagent is unstable and should be used the same day it is prepared; protect the Ellman's Reagent from light when not in use. Extra vials of the reagent have been provided should a plate need to be re-developed or multiple assays be run on different days.

2. Empty the wells and wash five times with Wash Buffer.
3. Add 200 µl of Ellman's Reagent to each well.
4. Add 5 µl of tracer to the TA wells.
5. Cover the plate with plastic film. Optimum development is obtained by using an orbital shaker equipped with a large, flat cover to allow the plate(s) to develop in the dark. This assay typically develops in 90-120 minutes. The longer the development the higher the O.D. values. If over-development occurs, wash the plate, add fresh Ellman's Reagent and allow to develop again.

Refer to page 28 for Procedure Summary.

Reading the Plate

1. Wipe the bottom of the plate with a clean tissue to remove fingerprints, dirt, etc.
2. Remove the plate cover being careful to keep Ellman's Reagent from splashing on the cover. *NOTE: Any loss of Ellman's Reagent will affect the absorbance readings. If Ellman's Reagent is present on the cover, use a pipette to transfer the Ellman's Reagent into the well. If too much Ellman's Reagent has splashed on the cover to easily redistribute back into the wells, wash the plate three times with Wash Buffer and repeat the development with fresh Ellman's Reagent.*
3. Read the plate at a wavelength between 405 and 420 nm.

ANALYSIS

Many plate readers come with data reduction software that plots data automatically. Alternatively a spreadsheet program can be used. The data should be plotted as either %B/B_0 *versus* log concentration using a four-parameter logistic fit or as logit B/B_0 *versus* log concentration using a linear fit. *NOTE: Cayman has a computer spreadsheet available for data anaylsis. Please contact Technical Service or visit our website (www.caymanchem.com/analysis/elisa) to obtain a free copy of this convenient data analysis tool.*

Calculations

Preparation of the Data

The following procedure is recommended for preparation of the data prior to graphical analysis.

NOTE: If the plate reader has not subtracted the absorbance readings of the blank wells from the absorbance readings of the rest of the plate, be sure to do that now.

1. Average the absorbance readings from the NSB wells.
2. Average the absorbance readings from the B_0 wells.
3. Subtract the NSB average from the B_0 average. This is the corrected B_0 or corrected maximum binding.
4. Calculate the B/B_0 (sample or standard bound/maximum bound) for the remaining wells. To do this, subtract the average NSB absorbance from the S1 absorbance and divide by the corrected B_0 (from Step 3). Repeat for S2-S8 and all sample wells. (To obtain %B/B_0 for a logistic four-parameter fit, multiply these values by 100.)

NOTE: The TA values are not used in the standard curve calculations. Rather, they are used as a diagnostic tool; the corrected B_0 divided by the actual TA (10X measured absorbance) will give the %Bound. This value should closely approximate the %Bound that can be calculated from the ***Representative Data*** *(see page 23). Erratic absorbance values and a low (or no) %Bound could indicate the presence of organic solvents in the buffer or other technical problems (see page 27 for* ***Troubleshooting****).*

Plot the Standard Curve

Plot %B/B_0 for standards S1-S8 *versus* MaR1 concentration using linear (y) and log (x) axes and perform a 4-parameter logistic fit.

Alternative Plot - The data can also be linearized using a logit transformation. The equation for this conversion is shown below. *NOTE: Do not use %B/B_0 in this calculation.*

$$\text{logit } (B/B_0) = \ln [B/B_0/(1 - B/B_0)]$$

Plot the data as logit (B/B_0) *versus* log concentrations and perform a linear regression fit.

Determine the Sample Concentration

Calculate the B/B_0 (or %B/B_0) value for each sample. Determine the concentration of each sample by identifying the %B/B_0 on the standard curve and reading the corresponding values on the x-axis. *NOTE: Remember to account for any concentration or dilution of the sample prior to the addition to the well.* Samples with %B/B_0 values greater than 80% or less than 20% should be re-assayed as they generally fall out of the linear range of the standard curve. A 20% or greater disparity between the apparent concentration of two different dilutions of the same sample indicates interference which could be eliminated by purification.

Performance Characteristics

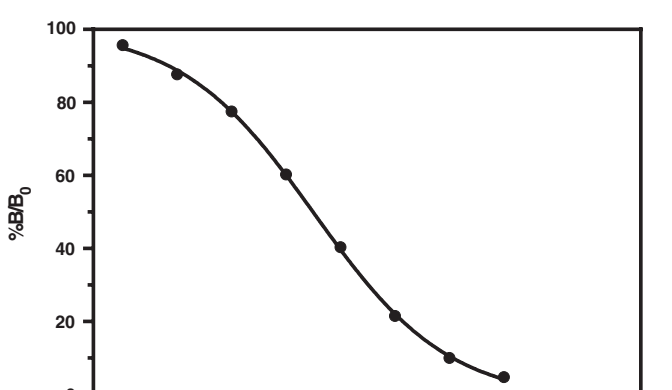
Representative Data

The standard curve presented here is an example of the data typically produced with this kit; however, your results will not be identical to these. You must run a new standard curve. Do not use the data below to determine the value of your samples.

Optical Density (O.D.) 414 nm at 90 min

MaR1 Standards (pg/ml)	Blank Corrected Absorbance	NSB Corrected Absorbance	%B/B_0	%CV Intra-Assay Precision	%CV Inter-Assay Precision
NSB	0.003	--	--	--	--
1,000	0.069	0.066	4.8	5.2	4.3
400	0.140	0.137	10.0	7.5	2.1
160	0.297	0.294	21.5	4.1	2.3
64	0.554	0.551	40.4	6.7	0.8
25.6	0.825	0.822	60.3	9.5	3.0
10.2	1.059	1.056	77.5	19.7	6.9
4.1	1.199	1.196	87.7	32.2*	18.0
1.6	1.308	1.305	95.7	41.8*	26.9

Table 2. Typical results
*Data in this range of the standard curve should be evaluated cautiously.



Assay Range = 1.64-1,000 pg/ml
Dynamic Range = 10.2-1,000 pg/ml
Sensitivity (defined as 80% B/B_0) = 9.6 pg/ml
Mid-point (defined as 50% B/B_0) = 30-70 pg/ml
Lower Limit of Detection = 5.8 pg/ml

The sensitivity and mid-point were derived from the standard curve shown above. The standard was diluted with ELISA Buffer.

Figure 6. Typical standard curve

Precision:

Intra-assay precision was determined by analyzing 24 replicates of three matrix controls (human urine samples) in a single assay.

Matrix Control (pg/ml)	%CV
132	4.6
62.8	5
16.3	5

Table 3. Intra-assay precision

Inter-assay precision was determined by analyzing replicates of three matrix controls (human urine samples) in separate assays spanning across several days.

Matrix Control (pg/ml)	%CV
200.1	10.0
39.9	14.8
12.8	28.1

Table 4. Inter-assay precision

Cross Reactivity:

Compound	Cross Reactivity
MaR1	100%
7-*epi* MaR1	46%
DHA	<0.01%
14S-HDHA	<0.01%
17R-HDHA	<0.01%
17S-HDHA	<0.01%
10S,17S-diHDHA	<0.01%
LTB_4	<0.01%
LXA_4	<0.01%
15-*epi* LXA_4	<0.01%
LXA_5	<0.01%
LXB_4	<0.01%
RvD1	<0.01%
17R-RvD1	<0.01%
RvD2	<0.01%
RvD3	<0.01%
RvE1	<0.01%

Table 5. Cross Reactivity of the MaR1 ELISA

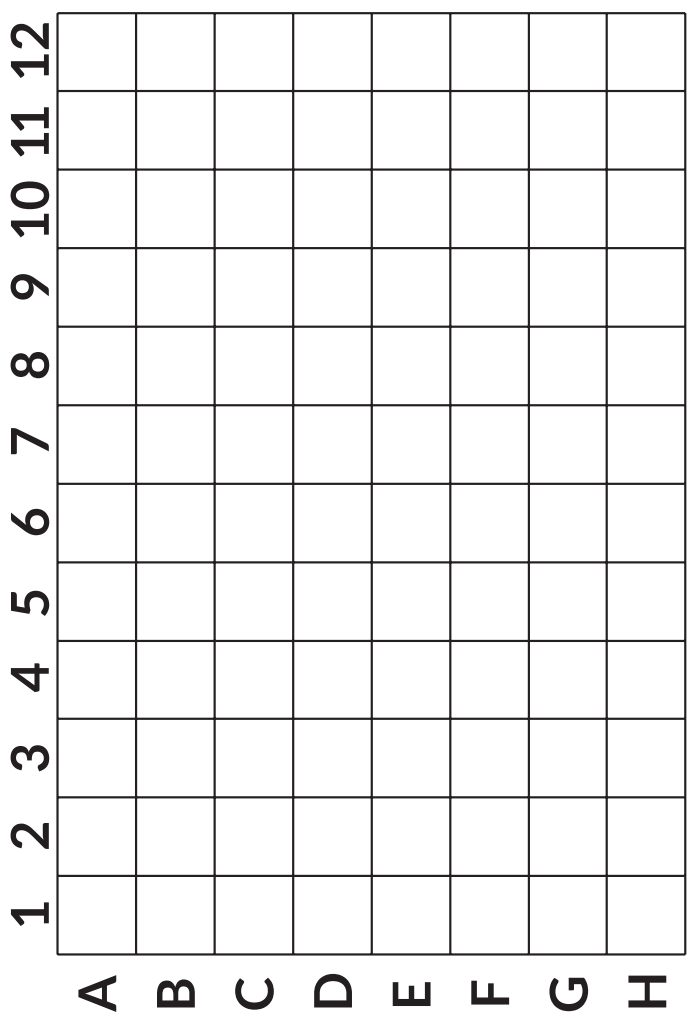
RESOURCES

Troubleshooting

Problem	Possible Causes	Recommended Solutions
Erratic values; dispersion of duplicates	A. Trace organic contaminants in the water source B. Poor pipetting/technique	A. Replace activated carbon filter or change source of UltraPure water
High NSB (>10% B_0)	A. Poor washing B. Exposure of NSB wells to specific antibody	A. Re-wash plate and redevelop
Very low B_0	A. Trace organic contaminants in the water source B. Plate requires additional development time C. Dilution error in preparing reagents	A. Replace activated carbon filter or change source of UltraPure water B. Return plate to shaker and re-read later
Low sensitivity (shift in dose response curve)	Standard is degraded	Replace standard
Analyses of two dilutions of a biological sample do not agree (*i.e.*, more than 20% difference)	Interfering substances are present	Purify sample prior to analysis by ELISA[5]
Only Total Activity (TA) wells develop	Trace organic contaminants in the water source	Replace activated carbon filter or change source of UltraPure water

Procedure	Blk	TA	NSB	B_0	Standards/Samples
Reconstitute and Mix	Mix all reagents gently				
ELISA Buffer (1X)	-	-	100 µl	50 µl	-
Standards/Samples	-	-	-	-	50 µl
Mar1 AChE Tracer	-	-	50 µl	50 µl	50 µl
Mar1 AChE Antiserum	-	-	-	50 µl	50 µl
Seal	Seal the plate and tap gently to mix				
Incubate	Incubate plate overnight at 4°C, static				
Aspirate	Aspirate wells and wash 5 x ~300 µl				
Apply Ellman's Reagent	200 µl	200 µl	200 µl	200 µl	200 µl
Total Activity (TA) - Tracer	-	5 µl	-	-	-
Seal	Seal plate and incubate 90-120 min at RT, shaking, protected from light				
Read	Read O.D. at 405-420 nm				

Table 6. MaR1 Assay Summary



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NOTES

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