

**ASSAY NAME: STI1\_BR**  
(Sexually transmitted infections panel 1 for BioRad)

**Quantity: 100 x 20µL PCR reactions**

**5-plex assay: *Chlamydia trachomatis*, *Haemophilus ducreyi*, *Mycoplasma hominis*, *Trichomonas vaginalis*, and human RPP30 DNA**

**SKU: PNP-STI1-D-BR-100 (Bio-Rad)**

**(RUO). Research Use Only. Not for use in Diagnostic Procedures.**

## SCOPE OF THIS PRODUCT INFORMATION SHEET (PIS):

The oligonucleotide recipes are optimized for each instrument (BioRad, QuantStudio, MIC). The verification data presented in this PIS were performed using PNP-STI1-D-BR-100 on a BioRad CFX96. The performance of the other SKUs on their corresponding instrument should be similar. Contact PCRassays.com if you need to use a different qPCR instrument.

## CONTENTS

The STI1 kit contains a mixture of primers/probes targeting the serine/threonine protein kinase (Protein ID: ATW18970.1) in the *Chlamydia trachomatis* genome, the gene of the ribonuclease r, virulence associated protein (Protein ID: VEG83041.1) in the *Haemophilus ducreyi* genome, the protein phosphatase 2C family (Protein ID: XP\_001315231.1) in the *Trichomonas vaginalis* genome, and the gene of the F0F1 ATP synthase subunit alpha (Protein ID: QHR23375.1) in the *Mycoplasma hominis* genome. The primers and probes in the STI1 assay are provided in Tube 1 as a 5X concentrated working solution that detects 4 pathogens and a human extraction control.

**Table of Dyes used in this assay:**

| Pathogen/Target       | Dyes          | Quencher | Refs. |
|-----------------------|---------------|----------|-------|
| <i>C. trachomatis</i> | <b>FAM</b>    | BHQ-1    | 1,2   |
| RPP30-DNA control     | <b>HEX</b>    | BHQ-1    | 7     |
| <i>H. ducreyi</i>     | <b>TEX615</b> | BHQ-2    | 3,4   |
| <i>T. vaginalis</i>   | <b>Cy5</b>    | BHQ-2    | 5     |
| <i>M. hominis</i>     | <b>Cy5.5</b>  | BHQ-2    | 6     |

The probes are designed as TaqMan<sup>®</sup> cleavage mechanism and thus the reaction requires a DNA polymerase with 5'-exonuclease activity.

## Assay contents:

**Tube 1:** Primer/Probe mix (5X) for *C. trachomatis*, *H. ducreyi*, *M. hominis*, *T. vaginalis*, and hRPP30DNA.

**Tube 2: (Do NOT add to specimen unknowns)** Positive control: 5000 copies/µl positive controls of synthetic 500 bp DNA fragments of *C. trachomatis*, *H. ducreyi*, *M. hominis*, *T. vaginalis*, and hRPP30DNA.

**Tube 3:** InhibiTaq Standard qPCR enzyme Mastermix (enough for 100 rxns. with 20 µL total volume). This is a custom formulation from Fortis Life Sciences to the specifications of PCRassays.com.



## ASSAY HANDLING

The STI1 assay is shipped at ambient temperature, but should be stored at -20 °C. The tubes should be kept on ice once thawed. Do not subject the enzyme to multiple freeze-thaw cycles.

Any contamination should be avoided by using appropriate personal protective equipment (PPE), powder free gloves, aerosol barrier pipette tips, and a clean hood.

## EXPERIMENTAL

Perform nucleic acid extraction/purification (recommended).

Set up your reaction (20 µL) as follows on ice:

| Component                            | Volume (µL) |
|--------------------------------------|-------------|
| InhibiTaq qPCR enzyme mastermix (2X) | 10          |
| Primer/Probe mix (5X)                | 4           |
| Sample                               | 2           |
| Water                                | 4           |

**Notes:** To improve assay sensitivity, up to 6 µL of sample can be added (water volume adjusted accordingly) for a total reaction volume of 20 µL. For positive control rxns., add 2 µL of the solution from Tube 2. Molecular biology grade water (NOT included) should be used to prepare the PCR reactions.

A PCR protocol was used for verification on a BioRad CFX96 system, with the following program:

| Step | Thermocycling Protocol:         |
|------|---------------------------------|
| 1    | Incubate @ 95 °C for 2 minutes  |
| 2    | Incubate @ 95 °C for 3 seconds  |
| 3    | Incubate @ 55 °C for 15 seconds |
| 4    | Plate Read                      |
| 5    | Go to Step 2, repeat 44× more   |

## RESULT INTERPRETATION

After running the qPCR reaction, perform a regression analysis on the data to determine the quantification cycle, Cq. (Cq is preferred over Ct). Each fluorescence channel with a Cq < 38 cycles and final RFU > 200 (BioRad) is considered “positive” or “+” in the Table below.

| <i>C. trachomatis</i><br>FAM™ | <i>H. ducreyi</i><br>TEX615™ | <i>T. vaginalis</i><br>Cy5™ | <i>M. hominis</i><br>Cy5.5™ | hRPP30<br>HEX™ | Recommended Interpretation                                                                                                                               |
|-------------------------------|------------------------------|-----------------------------|-----------------------------|----------------|----------------------------------------------------------------------------------------------------------------------------------------------------------|
| —                             | —                            | —                           | —                           | —              | The PCR reaction failed. Please repeat the experiment                                                                                                    |
| —                             | —                            | —                           | —                           | +              | The sample does not contain DNA of interest. The sample contains human RPP30 DNA.                                                                        |
| +                             | —                            | —                           | —                           | —              | The sample contains <i>C. trachomatis</i> DNA. The sample may not contain human RPP30 DNA.                                                               |
| +                             | —                            | —                           | —                           | +              | The sample contains <i>C. trachomatis</i> DNA and human RPP30 DNA.                                                                                       |
| —                             | +                            | —                           | —                           | —              | The sample contains <i>H. ducreyi</i> DNA. The sample may not contain human RPP30 DNA.                                                                   |
| —                             | +                            | —                           | —                           | +              | The sample contains <i>H. ducreyi</i> DNA and human RPP30 DNA.                                                                                           |
| —                             | —                            | +                           | —                           | —              | The sample contains <i>T. vaginalis</i> DNA. The sample may not contain human RPP30 DNA.                                                                 |
| —                             | —                            | +                           | —                           | +              | The sample contains <i>T. vaginalis</i> DNA and human RPP30 DNA.                                                                                         |
| —                             | —                            | —                           | +                           | —              | The sample contains <i>M. hominis</i> DNA. The sample may not contain human RPP30 DNA.                                                                   |
| —                             | —                            | —                           | +                           | +              | The sample contains <i>M. hominis</i> DNA and human RPP30 DNA.                                                                                           |
| +                             | +                            | +                           | +                           | —              | The sample contains <i>C. trachomatis</i> , <i>H. ducreyi</i> , <i>T. vaginalis</i> , <i>M. hominis</i> DNA. The sample may not contain human RPP30 DNA. |
| +                             | +                            | +                           | +                           | +              | The sample contains <i>C. trachomatis</i> , <i>H. ducreyi</i> , <i>T. vaginalis</i> , <i>M. hominis</i> DNA and human RPP30 DNA.                         |

## VERIFICATION EXPERIMENTS

The STI1 assay verification was carried out as a 5-plex assay that simultaneously detects DNA from *C. trachomatis*, *H. ducreyi*, *M. hominis*, *T. vaginalis*, and human RPP30 DNA, which serves as a positive control assay.

Experiments were performed in triplicate using the experimental procedure given above, but with different samples added to each reaction. The samples used for the verification experiments contained  $1 \times 10^4$  copies/reaction of synthetic 500 bp DNA constructs (from Twist Biosciences) harboring the regions of interest from the pathogen genomes, human RPP30 DNA gene, and human genomic DNA. **Figure 1** shows the results of these experiments, which indicate that the 5-plex specifically detects the different pathogens.

The limit of detection (LOD) was estimated by performing serial dilution experiments in triplicate (**Figure 2**). For dilution series only target construct was added. The results show a limit of detection (LOD) <10 copies/reaction.

## NOTES

<sup>1</sup> FAM™ (Carboxyfluorescein), a trademark of Life Technologies Corporation.

<sup>2</sup> BHQ-1™ (Black Hole Quencher) is a trademark of Biosearch Technologies, Inc.

<sup>3</sup> TEX615™ is a trademark of Thermo Fisher Scientific.

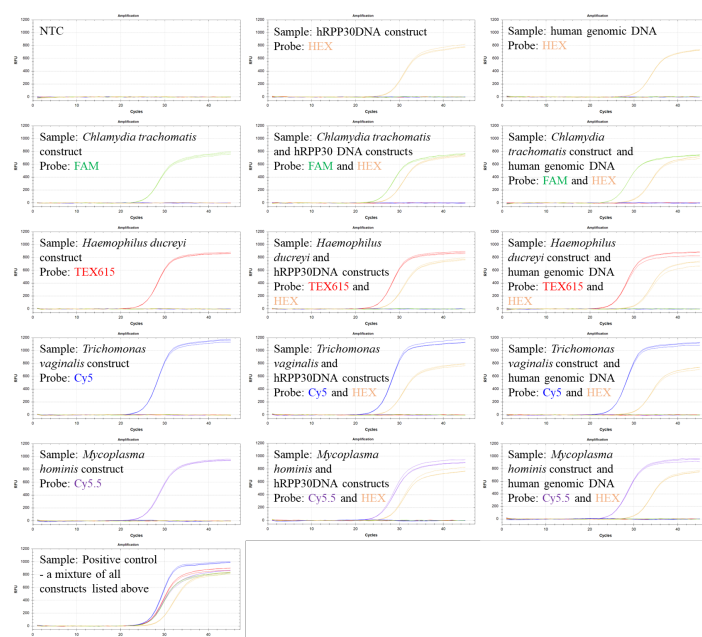
<sup>4</sup> BHQ-2™ (Black Hole Quencher) is a trademark of Biosearch Technologies, Inc.

<sup>5</sup> Cy5™, a trademark of GE Healthcare.

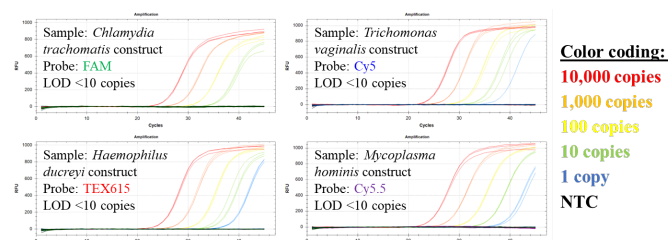
<sup>6</sup> Cy5.5™ is a trademark of GE Healthcare.

<sup>7</sup> HEX™ (Hexachloro-fluorescein), a trademark of Thermo Fisher Scientific.

<sup>8</sup> TaqMan™ is a trademark of Roche Diagnostics, Inc.



**Figure 1:** Verification experiments with single targets (given in text boxes for each panel). All sets of probes and primers are present in every reaction, but positive signal is only observed when the target(s) is present, indicating that the amplification is specific.



**Figure 2:** Serial dilution experiments show LOD <10 molecules for the synthetic DNA construct of each target.

**Conclusion:** The data in **Figure 1** indicate that the 5-plex primers and probes specifically detect and differentiate the pathogens and are also compatible with RPP30 DNA positive control primers. Human genomic DNA doesn't interfere with the detection of the pathogens.

## CONTACT US

For assistance, please contact DNA Software using the link:  
<https://www.pcrassays.com/contact/>

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