# S100A12 homodimer (human)

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# S100A12 homodimer (human)

# ELISA kit

# #A05084.96 wells

For research laboratory use only
Not for human diagnostic use

This assay has been developed
& validated by Bertin Bioreagent

Fabriqué en France
Made in France

#A11084

Version: 0121

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**96 wells**

**Storage: +4°C**

**Expiry date: stated on the package**

This kit contains:

| Designation | Colour of cap | Item # | Quantity per kit | Form |
|---|---|---|---|---|
| S100A12 homodimer (human) precoated 96-well Strip Plate | Blister with zip | A08084.1 ea | 1 | - |
| Streptavidin Poly_HRP Tracer | Green | A04410.100 dtn | 1 | Liquid |
| S100A12 homodimer (human) Biotin-labelled Antibody | Red | A03084.100 dtn | 1 | Lyophilised |
| S100A12 homodimer (human) Standard | Blue with red septum | A06084.1 ea | 2 | Lyophilised |
| S100A12 homodimer (human) Quality Control | Green with red septum | A010084.1ea | 2 | Lyophilised |
| Biotin-free ELISA Buffer | Grey / Blue | A07410.1 ea | 1 | Lyophilised |
| Wash Buffer | Silver | A17000.1 ea | 1 | Liquid |
| Tween 20 | Transparent | A12000.1 ea | 1 | Liquid |
| HRP Substrate Solution | Black | A09034.100 dtn | 1 | Liquid |
| HRP Stop Solution | Yellow | A22410.100 dtn | 1 | Liquid |
| Technical Booklet | - | A11084.1 ea | 1 | - |
| Well cover Sheet | - | - | 1 | - |

Each kit contains sufficient reagents for 96 wells. This allows for the construction of one standard curve in duplicate and the assay of 36 samples in duplicate. If you want to use the kit in two times, we provide one additional vial of Standard and one of Quality Control.

## Precaution for use

**Users are recommended to carefully read all instructions for use before starting work.**

Each time a new pipette tip is used, aspirate a sample or reagent and expel it back into the same vessel. Repeat this operation two or three times before distribution in order to equilibrate the pipette tip.

- For research laboratory use only
- Not for human diagnostic use
- Do not pipet liquids by mouth
- Do not use kit components beyond the expiration date
- Do not eat, drink or smoke in area in which kit reagents are handled
- Avoid splashing

HRP Stop Solution and HRP Substrate Solution are harmful solutions. To avoid any contact, wear eye, hand, face and clothing protection when handling these.

Wearing gloves, laboratory coat and glasses is recommended when assaying kit materials and samples.

### Temperature

Unless otherwise specified, all the experiments are done at room temperature (RT), that is around +20°C. Working at +25°C or more affects the assay and decreases its efficiency.

## Background

S100A12 belongs to the S100 family, called calgranulin. Calgranulins are endogenous molecules released in response to environmental triggers and cellular damage. Also known as Damage-Associated Molecular Pattern Molecules (DAMPs), these proteins play an important role in a diverse range of physiological and pathological processes, such as host defense, wound healing, autoimmunity, oncogenesis and inflammation, among others (Kerkhoff et al 2014; Holzinger et al 2018).

S100A12 is a calcium-, zinc- and copper- binding protein involved in processes that contribute to nutritional immunity against invading microbial pathogens. In humans, S100A12 (also named Calgranulin C or EN-RAGE) is mainly expressed and secreted by neutrophil granulocytes, and has been implicated in immune regulation. S100A12 is a ligand for the receptor for advanced glycation end products (RAGE), toll-like receptor 4 (TLR4), and CD36, which promote cellular and immunological pathways to alter inflammation (Garcia et al 2013).

S100A12 is co-expressed with two other S100 proteins, S100A8 (Mrp8) and S100A9 (Mrp14), the two subunits of

the calprotectin heterodimer, within granulocytes. S100A12 and S100A8/A9 proteins are encoded on the same chromosome, appear to be coregulated, and have functional and structural similarities (Oesterle & Bowman 2015). Given that S100A12 is intensely upregulated during trauma, infection, heat, stress and many other inflammatory processes, it is a valuable candidate as both a diagnostic biomarker and a therapeutic target for inflammation-associated diseases (Wang et al 2020). Plasma levels of S100A8/S100A9 and S100A12 were found to be higher in septic shock patients than in healthy volunteers. Furthermore, the high level of plasma calgranulins at admission in septic shock, were higher in non-survivors compared to survivors (Dubois et al 2019).

## Principle of the assay

This Enzyme Immunometric Assay (EIA/ELISA) is based on a sandwich technique. The wells of the plate are coated with a monoclonal antibody specific to S100A12 homodimer (human).

S100A12 homodimer (human) introduced into the wells (standard or sample) will be bound by the monoclonal antibody coated on the plate and is then detected by a second monoclonal antibody tagged with biotin also specific for S100A12 homodimer (human).

The two antibodies then form a sandwich by binding on different parts of the S100A12 homodimer (human).

The sandwich is immobilised on the plate so reagents in

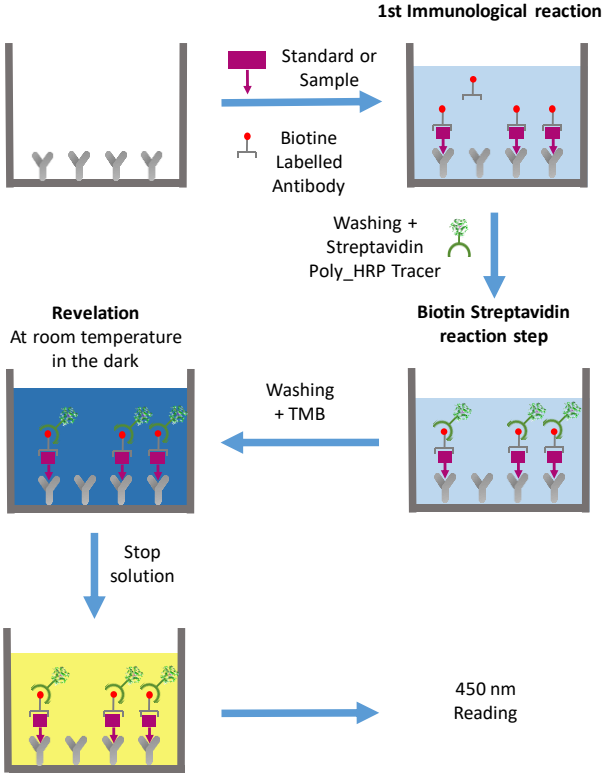
excess may be washed away.

The immunological complex is revealed by the interaction between biotin and streptavidin labelled with HRP (Tracer).

The concentration of S100A12 homodimer (human) is determined by measuring the enzymatic activity of immobilized Tracer using TMB. The Tracer acts on TMB to form a yellow compound after the reaction has been stopped.

The intensity of the colour, which is determined by spectrophotometry at 450 nm, is proportional to the amount of S100A12 homodimer (human) present in the well during the immunological incubation.

The principle of the assay is summarised below:



# Materials and equipment required

In addition to standard laboratory equipment, the following material is required:

For the assay:

- Precision micropipettes (20 to 1000 µL)
- Spectrophotometer plate reader (450 nm filter)
- Microplate washer (or washbottles)
- Orbital microplate shaker
- Multichannel pipette and disposable tips 30-300µL
- UltraPure water ELISA Grade#A07001
- Polypropylene tubes

Water used to prepare all ELISA reagents and buffers must be UltraPure ELISA Grade (deionized & free from organic contaminant traces).
Do not use distilled water, HPLC-grade water or sterile water.

- UltraPure water ELISA Grade may be purchased from Bertin Bioreagent (item #A07001).

The shaking speed is an important parameter and it must vigorous enough to agitate well content and without splashing droplets into the other wells (around 500 rpm, speed to be adapter do the device)

# Sample type, collection and preparation

This assay has been validated to measure S100A12 homodimer (human) in human serum.

It is the responsibility of the user to check the compatibility of the assay with the study matrix.

## General precautions

- All samples must be free from organic solvents prior to assay.
- Samples should be assayed immediately after collection or should be stored at -20°C or at -80°C prior the use with the assay.

## Sample collection

Blood samples are collected in dry tubes.

## Sample preparation

Serum sample must be diluted at least at 1:5 in 1x Biotin-free ELISA Buffer before the assay. The dilution of highly concentrated samples should be optimized by the user.

# Reagent preparation

Each kit contains sufficient reagents for 96 wells. This allows for the construction of one standard curve in duplicate and

the assay of 36 samples in duplicate.

If you want to use the kit in two times, we provide one additional vial of Standard and one of Quality Control.

All reagents need to be brought to room temperature (around +20°C) prior to the assay.

### 1x Biotin-free ELISA Buffer

Reconstitute the Biotin-free ELISA Buffer #A07084 with 50 mL of UltraPure water ELISA Grade . Allow it to stand 5 minutes until completely dissolved and then mix thoroughly by gentle inversion.
*Stability at 4°C: 3 weeks.*

### S100A12 homodimer (human) Standard

Reconstitute one S100A12 homodimer (human) Standard vial #A06084 with 1 mL of UltraPure water ELISA Grade. Allow it to stand for 5 minutes until completely dissolved and then mix thoroughly by gentle inversion.

The concentration of the first standard (S1) is 400.0 ng/mL. Prepare seven polypropylene tubes (for the seven other standards) and add 500 µL of 1x Biotin-free ELISA Buffer into each tube. Then prepare the standards by serial dilutions as follow:

| Standard | Volume of Standard | Volume of 1 X Biotin-free ELISA Buffer | Standard concentration |
|---|---|---|---|
| S1 | - | - | 400.0 ng/mL |
| S2 | 500 µL of S1 | 500 µL | 200.0 ng/mL |
| S3 | 500 µL of S2 | 500 µL | 100.0 ng/mL |
| S4 | 500 µL of S3 | 500 µL | 50.0 ng/mL |
| S5 | 500 µL of S4 | 500 µL | 25.0 ng/mL |
| S6 | 500 µL of S5 | 500 µL | 12.5 ng/mL |
| S7 | 500 µL of S6 | 500 µL | 6.3 ng/mL |
| S8 | 500 µL of S7 | 500 µL | 3.1 ng/mL |

*Stability at 4°C: With in the day.*

### S100A12 homodimer (human) Quality Control

Reconstitute one S100A12 homodimer (human) QC vial #A10084 with 1 mL of UltraPure water ELISA Grade. Allow it to stand 5 minutes until completely dissolved and then mix thoroughly by gentle inversion.
*Stability at 4°C: With in the day.*

### S100A12 homodimer (human) Biotin-labelled Antibody

Reconstitute the S100A12 homodimer (human) Biotin-labelled Antibody vial #A03084 with 5 mL of 1x Biotin-free

ELISA Buffer. Allow it to stand 5 minutes until completely dissolved and then mix thoroughly by gentle inversion.
*Stability of diluted antibody at +4°C: 3 weeks.*

### Wash Buffer

Dilute 2 mL of concentrated Wash Buffer #A17000 with 800 mL of UltraPure water ELISA Grade. Add 400 µL of Tween 20 #A12000. Use a magnetic stirring bar to mix the content.
*Stability at +4°C: 3 weeks.*

## Assay procedure

It is recommended to perform the assays in duplicate following the instructions hereafter.

### Plate preparation

Prepare the Wash Buffer as indicated in the reagent preparation section.

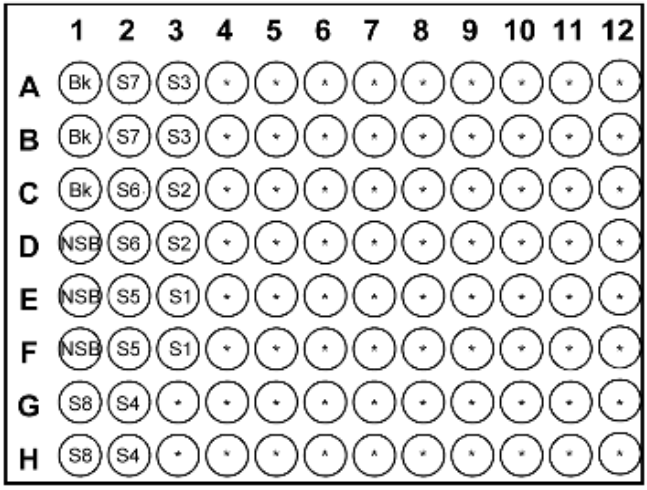
Open the plate pouch and select enough strips for your assay and place the unused strips back in the pouch.
*Stability at +4°C: 1 month.*

Rinse each well 5 times with Wash Buffer (300 µL/well).

Just before distributing the reagents and samples, remove the buffer from the wells by inverting the plate and shaking out the last drops on a paper towel.

## Plate set-up

A plate set-up is suggested hereafter. The contents of each well may be recorded on the template sheet provided at the end of this technical booklet.




Bk : Blank
NSB : Non Specific Binding
S1-S8 : Standards 1-8
* : Samples or Quality Controls

## Pipetting the reagents

All samples and reagents must reach room temperature prior to performing the assay.
Use different tips to pipet the buffers, standards, samples, antibody and other reagents.

Before pipetting, equilibrate the pipette tips in each reagent. Do not touch the liquid already in the well when expelling with the pipette tip.

- ***1x Biotin-free ELISA Buffer***

Dispense 50 µL to Non Specific Binding wells (NSB) wells and 40 µl in samples wells.

- ***S100A12 homodimer (human) Standard and QC***

Dispense 50 µL of each of the eight standards (S8 to S1) and QC in duplicate to appropriate wells.

Start with the lowest concentration standard (S8) and equilibrate the tip in the next higher standard before pipetting.

- ***S100A12 homodimer samples***

Dispense 10 µL in duplicate to appropriate wells. See section "Sample type, collection and preparation" for dilution of samples.

- ***S100A12 homodimer (human) Biotin-labelled antibody***

Dispense 50 µL to each well, except Blank (Bk) wells.

## Incubating the plate

Cover the plate with the cover sheet and incubate 60 minutes at room temperature, shaking on an orbital microplate shaker (See shaking speed advices page 11).

### Washing the plate

Rinse each well 5 times with Wash Buffer (300 µL/well). Just before distributing reagents, remove the buffer from the wells by inverting the plate and shaking out the last drops on a paper towel.

### Pipetting the reagents

> ***Streptavidin Poly_HRP Tracer***

Dispense 50 µL to each well, except Blank (Bk) wells.

### Incubating the plate

Cover the plate with the cover sheet and incubate 30 minutes at room temperature, shaking on an orbital microplate shaker (>See shaking speed advices page 11).

### Developing and reading the plate

- Empty the plate by turning it over. Rinse each well 5 times with 300 µL of Wash Buffer. At the end of the last washing step, empty the plate and blot the plate on a paper towel to discard any trace of liquid.

- Add 100µL of HRP Substrate Solution to each well.

- Incubate the plate in the dark at room temperature **without shaking**. ***To set up the incubation time, please refer to the Quality Control Sheet (QCS) corresponding to the lot#***.

- Add 100µL of HRP Stop Solution to each well.
- Wipe the bottom of the plate with a paper towel, and make sure that no liquid has splashed outside the wells.
- Read the plate at 450 nm (yellow color).

## Assay procedure summary

| **Enzyme Immunoassay Protocol (volumes are in µL)** | | | | |
|---|---|---|---|---|
| | Blank | NSB | Standard Or QC | Sample |
| Biotin-free ELISA Buffer | - | 50 | - | 40 |
| Standard | - | - | 50 | - |
| Sample | - | - | - | 10 |
| Biotin-Labelled Antibody | - | 50 | | |
| Cover plate, incubate **60** minutes at room temperature under orbital shaking* | | | | |
| Wash strips 5 times with 300 µL/well Discard liquid from the wells & dry on absorbent paper - | | | | |
| Streptavidin Poly_HRP Tracer | - | 100 | | |
| Cover plate, incubate **30** minutes at room temperature under orbital shaking* | | | | |
| Wash strips 5 times with 300 µL/well Discard liquid from the wells & dry on absorbent paper | | | | |
| HRP Substrate Solution | 100 | | | |
| Incubate the plate in the dark without agitation | | | | |
| HRP Stop Solution | 100 | | | |
| Read the plate at 450 nm | | | | |

*The shaking speed is an important parameter and it must vigorous enough to agitate well content and without splashing droplets into the other wells (around 500 rpm, speed to be adapter do the device).

# Data analysis

Make sure that your plate reader has subtracted the absorbance readings of the blank wells from the absorbance readings of the rest of the plate. If not, do it at this steps.

- Calculate the average absorbance for each NSB, standard, QC and sample.
- For each standard, plot the absorbance on y axis versus the concentration on x axis. Draw a best-fit line through the points.
- To determine the concentration of your samples, find the absorbance value of each sample on the y axis.
- Read the corresponding value on the x axis which is the concentration of your unknown sample.
- Samples with a concentration greater than 400.00 ng/mL should be re-assayed after dilution in Biotin-free ELISA Buffer.
- Most plate readers are supplied with a curve-fitting software capable of graphing these data (4-parameter logistic fit 4PL). If you have this type of software, we recommend using it. Refer to it for further information.

**Two vials of Quality Control are provided with this kit.**
**Your standard curve is validated only if the calculated concentration of the Quality Control obtained with the assay is +/- 25% of the expected concentration (see the label of the QC vial)**

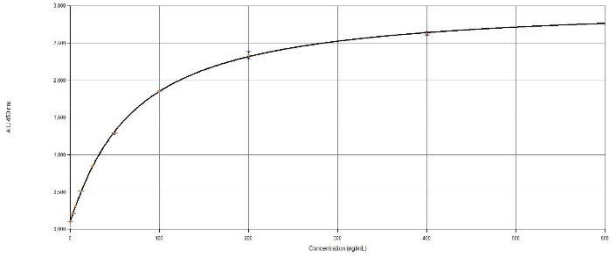
## Acceptable range

- NSB absorbance ≤ 0.150 A.U.
- Limit of detection ≤ 2.0 ng/mL
- QC ±25% of the expected concentration (see the label of the QC vial)
- Absorbance values (blank-deducted) of the standard S1 ≥ 2.2
- Ratio:[Absorbance values (blank-deducted) of the standard S8]/[Absorbance values (blank-deducted) of the NSB] ≥ 1.5
- Curve analysis method:
  - recommended: 4 parameter logistic fitting (4PL)
  - Alternative 4 parameter logistic fitting with a 1/Y ponderation (4-PL 1/Y)

## Typical results

The following data are for demonstration purpose only. Your data may be different and still correct.
These data were obtained using all reagents as supplied in this kit under the following conditions: 20 minutes developing at room temperature, reading at 450 nm. A 4 parameter logistic fitting was used to determine the concentrations.

| Standard | S100A12 homodimer (human) ng/mL | Absorbance A.U. |
|---|---|---|
| S1 | 400.0 | 2.630 |
| S2 | 200.0 | 2.338 |
| S3 | 100.0 | 1.853 |
| S4 | 50.0 | 1.288 |
| S5 | 25.0 | 0.852 |
| S6 | 12.5 | 0.513 |
| S7 | 6.3 | 0.312 |
| S8 | 3.1 | 0.215 |
| NSB | 0.0 | 0.100 |



Typical S100A12 homodimer (human) standard curve

# Assay characteristics

- ***Validated for***
  - Human serum samples
- ***Limit of detection (LOD):*** *≤2.0 ng/mL (calculated as the concentration of* S100A12 homodimer (human) *corresponding to the NSB average plus three standard deviations)*
- ***Cross-reactivity***

| Molecule/Species | Cross-reactivity |
|---|---|
| S100A8/S100A9 (human) | Not detected |

- ***Inter-assay variation***

| QC level | QC 1 | QC 2 | QC 3 |
|---|---|---|---|
| Mean concentration (ng/mL) | 34.4 | 84.3 | 276.3 |
| Mean concentration (ng/mL) x dilution | 172.2 | 421.6 | 1381.4 |
| CV % | 7.4 | 8.8 | 22.6 |

QC are sera spiked or un-spiked with S100A12 homodimer (human) and them diluted at 1:5 (10 µl QC + 40 µL of 1x

Biotin-free ELISA Buffer) before testing.

For the inter–assay validation, the number of replicates is equal to 6 for each level of QC. The three validation level were analyzed along with the calibration curve for a total of 6 independent runs.

> ***Intra-assay variation***

| QC level | QC 1 | QC 2 | QC 3 |
|---|---|---|---|
| Mean concentration (ng/mL) | 35.0 | 83.3 | 251.7 |
| Mean concentration (ng/mL) x dilution | 175.0 | 416.6 | 1258.6 |
| CV % | 4.9 | 6.3 | 10.0 |

QC are sera spiked or un-spiked with S100A12 homodimer (human) and them diluted at 1:5 (10 µl QC + 40 µL of 1x Biotin-free ELISA Buffer) before testing.

For the intra–assay validation, the number of replicates is equal to 10 for each level of QC. The three validation level were analyzed along with the calibration curve for a unique experiments.

### > *Matrix variability*

| Matrix# | Mean endogenous conc.. x dil.. factor (1) (ng/mL) | Mean spiked matrix conc.. x dil.. factor (1) (ng/mL) | Back calculated conc.. (ng/mL) | Back calculated spiked conc.. (Relative error) |
|---|---|---|---|---|
| 1 | 140.7 | 389.7 | 249.1 | 9.4 % |
| 2 | 150.9 | 422.3 | 271.4 | 19.2 % |
| 3 | 383.7 | 691.5 | 307.7 | 30.4 % |
| 4 | 378.3 | 692.1 | 313.8 | 32.9 % |
| 5 | 864.1 | 1128.0 | 263.9 | 11.8 % |
| 6 | 427.8 | 666.8 | 239.0 | 1.3 % |
| 7 | 662.8 | 900.5 | 237.7 | - 4.9 % |
| 8 | 184.7 | 446.4 | 261.7 | 4.7 % |
| 9 | 234.8 | 484.0 | 249.2 | - 0.3 % |

(1) Matrix dilution is 1:5

9 individual sera samples were spiked and not with spiked with S100A12 homodimer (human). Each sample (spiked or not) was evaluated 5 fold diluted in duplicate and analyzed against a calibration curve.

### > *Linearity*

Three individual sera were spiked or not with S100A12 homodimer (human) and diluted at 1:5
Each spiked diluted sample is evaluated on 5 serial dilutions

| Matrix# | Endogenous Conc. (ng/mL) | Dilution | Spiked S100A12 homodimer (human ) (ng/mL) | Mean of spiked conc. x dil. (ng/mL) | CV % Mean of spiked conc. x dil. | Recovery % | Mean Recovery % |
|---|---|---|---|---|---|---|---|
| 1 | 8.7 | 1 | 256.6 | 252.6 | 11.1 | 89.0 | 111.8 |
| | | 2 | 171.7 | 343.4 | | 121.0 | |
| | | 4 | 86.0 | 343.8 | | 121.1 | |
| | | 8 | 42.0 | 336.3 | | 118.5 | |
| | | 16 | 20.3 | 325.6 | | 114.7 | |
| | | 32 | 9.5 | 302.6 | | 106.6 | |
| 2 | 24.3 | 1 | 241.5 | 241.5 | 15.4 | 80.6 | 113.7 |
| | | 2 | 175.1 | 350.2 | | 116.9 | |
| | | 4 | 96.1 | 384.5 | | 128.4 | |
| | | 8 | 47.7 | 381.6 | | 127.4 | |
| | | 16 | 22.1 | 353.0 | | 117.9 | |
| | | 32 | 10.4 | 332.7 | | 111.1 | |

| Matrix# | Endogenous Conc. (ng/mL) | Dilution | Spiked S100A12 homodimer (human ) (ng/mL) | Mean of spiked conc. x dil. (ng/mL) | CV % Mean of spiked conc. x dil. | Recovery % | Mean Recovery % |
|---|---|---|---|---|---|---|---|
| 3 | 144.8 | 1 | 312.0 | 312.0 | 17.4 | 74.3 | 109.9 |
| | | 2 | 249.5 | 499.1 | | 118.9 | |
| | | 4 | 135.2 | 540.6 | | 128.8 | |
| | | 8 | 62.2 | 497.5 | | 118.5 | |
| | | 16 | 30.0 | 479.6 | | 114.2 | |
| | | 32 | 13.8 | 440.5 | | 104.9 | |

### > *Parallellism*

Five individual human sera samples were diluted between 1:5 and 1:80 by serial dilution. Each dilution was analysed against a calibration curve.

| Matrix# | Dilution factor | S100A12 homodimer (human) conc. (ng/mL) | S100A12 homodimer (human) conc. (ng/mL) X Dilution factor | CV% |
|---|---|---|---|---|
| 1 | 5 | 72.5 | 362.5 | 9.3 % |
| | 10 | 38.4 | 383.8 | |
| | 20 | 20.2 | 403.5 | |
| | 40 | 8.6 | 344.6 | |
| | 80 | 4.0 | 316.7 | |
| 2 | 5 | 162.9 | 814.6 | 7.8 % |
| | 10 | 91.3 | 913.2 | |
| | 20 | 45.9 | 918.6 | |
| | 40 | 20.9 | 836.4 | |
| | 80 | 9.5 | 762.8 | |
| 3 | 5 | 130.5 | 652.5 | 5.1 % |
| | 10 | 66.1 | 661.0 | |
| | 20 | 33.7 | 673.5 | |
| | 40 | 15.8 | 631.7 | |
| | 80 | 7.4 | 590.0 | |
| 4 | 5 | 36.8 | 183.9 | 8.0 % |
| | 10 | 19.6 | 196.4 | |
| | 20 | 10.0 | 200.2 | |
| | 40 | 4.7 | 186.8 | |
| | 80 | 2.0 | 162.3 | |
| 5 | 5 | 43.6 | 218.0 | 7.8 % |
| | 10 | 23.2 | 231.9 | |
| | 20 | 12.3 | 246.1 | |
| | 40 | 5.4 | 214.4 | |
| | 80 | 2.5 | 201.0 | |

# Troubleshooting

### *Absorbance values are too low:*

- one reagent has not been dispensed,
- too low shaking
- incorrect preparation,
- assay performed before reagents reached room temperature,
- reading time not long enough.

### *High signal and background in all wells:*

- inefficient washing,
- overdeveloping (incubation time should be reduced),
- high ambient temperature.

### *High dispersion of duplicates:*

- poor pipetting technique
- irregular plate washing.

These are a few examples of troubleshooting that may occur. If you need further explanation, Bertin Bioreagent will be happy to assist you. Feel free to contact our Technical Support staff by phone +33 (0)139 306 036 or E-mail tech@bertin-bioreagent.com, and be sure to indicate the batch number of the kit (see outside the box).

Bertin Bioreagent proposes ELISA Training kit #B05005. For further information, please contact our Technical Support.

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We strive to address a broad range of research interest: inflammation, oxidative injury, endocrinology, diabetes, obesity, hypertension, pain, prion diseases.

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# CONTACT US

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