Cysteinyl Leukotriene Express ELISA Kit

Item No. 10009291

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Customer Service 800.364.9897
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1180 E. Ellsworth Rd · Ann Arbor, MI · USA

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GENERAL INFORMATION

Materials Supplied

Item Number	Item	96 wells Quantity/Size	480 wells Quantity/Size
10011322	Cysteinyl Leukotriene Express ELISA Monoclonal Antibody	1 vial/100 dtn	1 vial/500 dtn
10011323	Cysteinyl Leukotriene Express AChE Tracer	1 vial/100 dtn	2 vials/500 dtn
420504	Cysteinyl Leukotriene Express ELISA Standard	1 vial	1 vial
10011325	Immunoassay Buffer A Concentrate (10X)	2 vials/10 ml	4 vials/10 ml
400062	Wash Buffer Concentrate (400X)	1 vial/5 ml	1 vial/12.5 ml
400035	Polysorbate 20	1 vial/3 ml	1 vial/3 ml
400008/400009	Goat Anti-Mouse IgG Coated Plate	1 plate	5 plates
400012	96-Well Cover Sheet	1 cover	5 covers
400050	Ellman's Reagent	3 vials/100 dtn	6 vials/250 dtn

If any of the items listed above are damaged or missing, please contact our Customer Service department at (800) 364-9897 or (734) 971-3335. We cannot accept any returns without prior authorization.

WARNING: THIS PRODUCT IS FOR RESEARCH ONLY - NOT FOR HUMAN OR VETERINARY DIAGNOSTIC OR THERAPEUTIC USE.

Safety Data

This material should be considered hazardous until further information becomes available. Do not ingest, inhale, get in eyes, on skin, or on clothing. Wash thoroughly after handling. Before use, the user must review the complete Safety Data Sheet, which has been sent *via* email to your institution.

Precautions

Please read these instructions carefully before beginning this assay.

The reagents in this kit have been tested and formulated to work exclusively with Cayman Chemical's Cysteinyl Leukotriene Express ELISA Kit. This kit may not perform as described if any reagent or procedure is replaced or modified.

When compared to quantification by LC/MS or GC/MS, it is not uncommon for immunoassays to report higher analyte concentrations. While LC/MS or GC/MS analyses typically measure only a single compound, antibodies used in immunoassays sometimes recognize not only the target molecule, but also structurally related molecules, including biologically relevant metabolites. In many cases, measurement of both the parent molecule and metabolites is more representative of the overall biological response than is the measurement of a short-lived parent molecule. It is the responsibility of the researcher to understand the limits of both assay systems and to interpret their data accordingly.

If You Have Problems

Technical Service Contact Information

Phone: 888-526-5351 (USA and Canada only) or 734-975-3888

Email: techserv@caymanchem.com

In order for our staff to assist you quickly and efficiently, please be ready to supply the lot number of the kit (found on the outside of the box).

Storage and Stability

This kit will perform as specified if stored as directed at -80°C and used before the expiration date indicated on the outside of the box.

Materials Needed But Not Supplied

1. A plate reader capable of measuring absorbance between 405-420 nm.
2. Adjustable pipettes and a repeating pipettor.
3. A source of 'UltraPure' water. Water used to prepare all ELISA reagents and buffers must be deionized and free of trace organic contaminants ('UltraPure'). Use activated carbon filter cartridges or other organic scavengers. Glass distilled water (even if double distilled), HPLC-grade water, and sterile water (for injections) are not adequate for ELISA. *NOTE: UltraPure water is available for purchase from Cayman (Item No. 400000).*

INTRODUCTION

Background

The leukotrienes (LTs) were discovered in 1979 as a group of acute inflammatory mediators derived from arachidonic acid in leukocytes.[1,2] Their biosynthesis was shown to proceed *via* the 5-lipoxygenase (5-LO) pathway. LT biosynthesis has subsequently been demonstrated in other bone marrow-derived cells expressing 5-LO including eosinophils, mast cells, and macrophages.

5-LO converts arachidonic acid into LTA_4 with 5(S)-HpETE as an intermediate. The conjugation of glutathione to LTA_4 results in the formation of LTC_4. LTC_4 is rapidly metabolized to LTD_4 and LTE_4 as shown in Figure 1 (see page 7).[3] This metabolism is essentially complete within 10 minutes in the human lung. LTC_4, LTD_4, and LTE_4 are collectively referred to as cysteinyl leukotrienes (CysLTs).

LTC_4 and LTD_4 are potent mediators of asthma and hypersensitivity. They induce bronchoconstriction, increase microvascular permeability, and are vasoconstrictors of coronary arteries.[4] The biological activity of LTE_4 is much lower in most systems studied, but its presence reflects the prior existence of LTC_4 and LTD_4.

About This Assay

Cayman's CysLT Express ELISA Kit is a competitive assay that can be used for quantification of CysLT in urine, culture media, and other sample matrices. The assay has a range of 7.8-1,000 pg/ml and a sensitivity (80% B/B_0) of approximately 20 pg/ml.

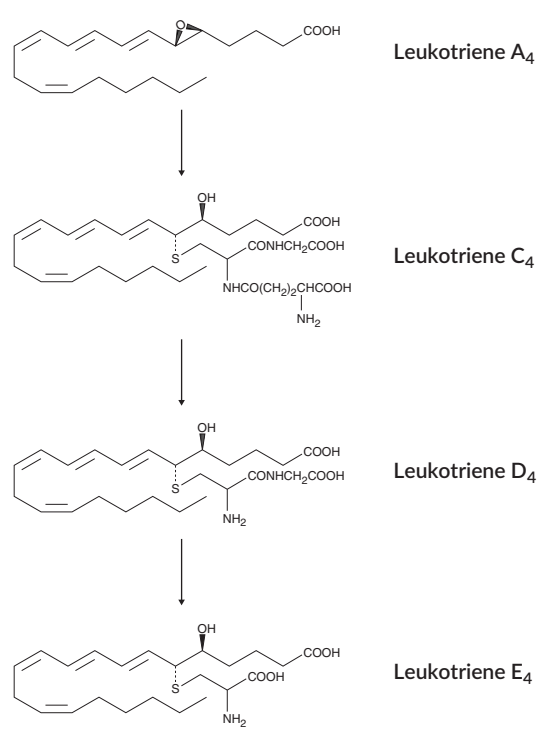


Figure 1. The formation of CysLTs

Description of AChE Competitive ELISAs[5,6]

This assay is based on the competition between CysLTs and a CysLT-acetylcholinesterase (AChE) conjugate (CysLT Tracer) for a limited amount of CysLT ELISA Monoclonal Antibody. Because the concentration of the CysLT Tracer is held constant while the concentration of CysLT varies, the amount of CysLT Tracer that is able to bind to the CysLT Monoclonal Antibody will be inversely proportional to the concentration of CysLT in the well. This antibody-CysLT complex binds to a goat polyclonal anti-mouse IgG that has been previously attached to the well. The plate is washed to remove any unbound reagents and then Ellman's Reagent (which contains the substrate to AChE) is added to the well. The product of this enzymatic reaction has a distinct yellow color and absorbs strongly at 412 nm. The intensity of this color, determined spectrophotometrically, is proportional to the amount of CysLT Tracer bound to the well, which is inversely proportional to the amount of free CysLT present in the well during the incubation; or

$$\text{Absorbance} \propto \text{[Bound CysLT Tracer]} \propto 1/\text{[CysLT]}$$

A schematic of this process is shown in Figure 2, on page 9.

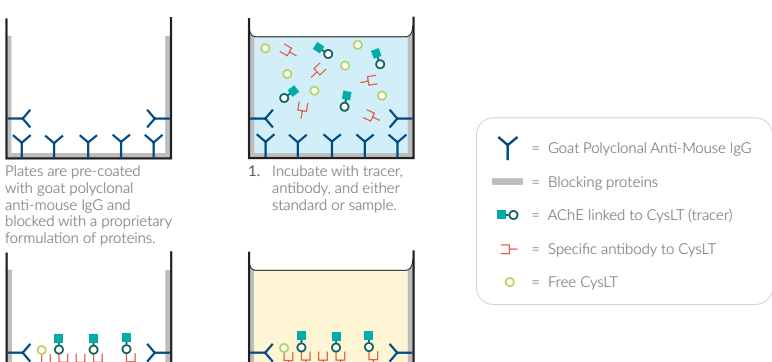


Figure 2. Schematic of the AChE ELISA

Biochemistry of Acetylcholinesterase

The electric organ of the electric eel, *E. electricus*, contains an avid AChE capable of massive catalytic turnover during the generation of its electrochemical discharges. The electric eel AChE has a clover leaf-shaped tertiary structure consisting of a triad of tetramers attached to a collagen-like structural fibril. This stable enzyme is capable of high turnover (64,000 s^{-1}) for the hydrolysis of acetylthiocholine.

A molecule of the analyte covalently attached to a molecule of AChE serves as the tracer in AChE enzyme immunoassays. Quantification of the tracer is achieved by measuring its AChE activity with Ellman's Reagent. This reagent consists of acetylthiocholine and 5,5'-dithio-*bis*-(2-nitrobenzoic acid). Hydrolysis of acetylthiocholine by AChE produces thiocholine (see Figure 3, on page 11). The non-enzymatic reaction of thiocholine with 5,5'-dithio-*bis*-(2-nitrobenzoic acid) produces 5-thio-2-nitrobenzoic acid, which has a strong absorbance at 412 nm (ε = 13,600).

AChE has several advantages over other enzymes commonly used for enzyme immunoassays. Unlike horseradish peroxidase, AChE does not self-inactivate during turnover. This property of AChE also allows redevelopment of the assay if it is accidentally splashed or spilled. In addition, the enzyme is highly stable under the assay conditions, has a wide pH range (pH 5-10), and is not inhibited by common buffer salts or preservatives. Since AChE is stable during the development step, it is unnecessary to use a 'stop' reagent, and the plate may be read whenever it is convenient.

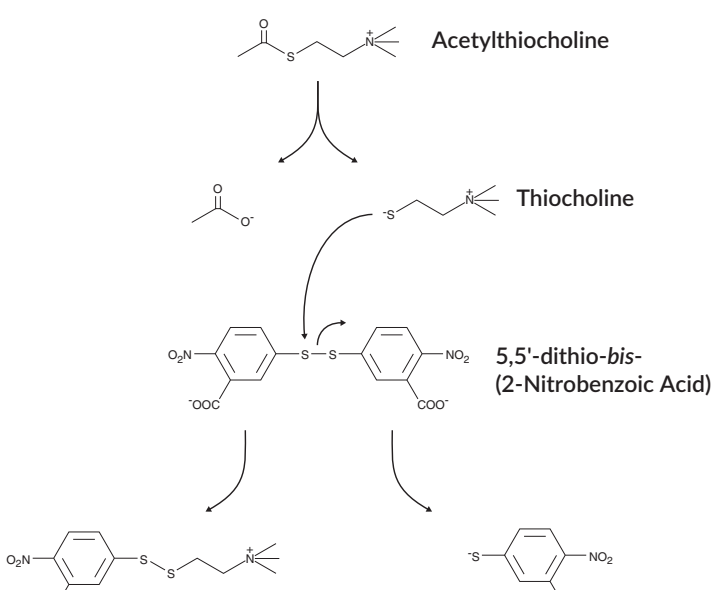


Figure 3. Reaction catalyzed by acetylcholinesterase

Definition of Key Terms

Blank: background absorbance caused by Ellman's Reagent. The blank absorbance should be subtracted from the absorbance readings of all the other wells, including NSB wells.

Total Activity: total enzymatic activity of the AChE-linked tracer. This is analogous to the specific activity of a radioactive tracer.

NSB (Non-Specific Binding): non-immunological binding of the tracer to the well. Even in the absence of specific antibody a very small amount of tracer still binds to the well; the NSB is a measure of this low binding. Do not forget to subtract the Blank absorbance values.

B_0 (Maximum Binding): maximum amount of the tracer that the antibody can bind in the absence of free analyte.

%B/B_0 (%Bound/Maximum Bound): ratio of the absorbance of a particular sample or standard well to that of the maximum binding (B_0) well.

Standard Curve: a plot of the %B/B_0 values *versus* concentration of a series of wells containing various known amounts of analyte.

Dtn: determination, where one dtn is the amount of reagent used per well.

Cross Reactivity: numerical representation of the relative reactivity of this assay towards structurally related molecules as compared to the primary analyte of interest. Biomolecules that possess similar epitopes to the analyte can compete with the assay tracer for binding to the primary antibody. Substances that are superior to the analyte in displacing the tracer result in a cross reactivity that is greater than 100%. Substances that are inferior to the primary analyte in displacing the tracer result in a cross reactivity that is less than 100%. Cross reactivity is calculated by comparing the mid-point (50% B/B_0) value of the tested molecule to the mid-point (50% B/B_0) value of the primary analyte when each is measured in assay buffer using the following formula:

$$\%\ \text{Cross Reactivity} = \left[\frac{50\%\ B/B_0 \text{ value for the primary analyte}}{50\%\ B/B_0 \text{ value for the potential cross reactant}}\right] \times 100\%$$

PRE-ASSAY PREPARATION

NOTE: Water used to prepare all ELISA reagents and buffers must be deionized and free of trace organic contaminants ('UltraPure'). Use activated carbon filter cartridges or other organic scavengers. Glass distilled water (even if double distilled), HPLC-grade water, and sterile water (for injections) are not adequate for ELISA. UltraPure water may be purchased from Cayman (Item No. 400000).

Buffer Preparation

Store all diluted buffers at 4°C; they will be stable for about two months.

1. **Immunoassay Buffer A Preparation**

   Dilute the contents of one vial of Immunoassay Buffer A Concentrate (10X) (Item No. 10011325) with 90 ml of UltraPure water. Be certain to rinse the vial to remove any salts that may have precipitated. *NOTE: It is normal for the concentrated buffer to contain crystalline salts after thawing. These will completely dissolve upon dilution with water.*

2. **Wash Buffer Preparation**

   5 ml vial Wash Buffer Concentrate (400X) (96-well kit; Item No. 400062): Dilute to a total volume of 2 liters with UltraPure water and add 1 ml of Polysorbate 20 (Item No. 400035).

   OR

   12.5 ml vial Wash Buffer Concentrate (400X) (480-well kit; Item No. 400062): Dilute to a total volume of 5 liters with UltraPure water and add 2.5 ml of Polysorbate 20 (Item No. 400035).

Smaller volumes of Wash Buffer can be prepared by diluting the Wash Buffer Concentrate 1:400 and adding Polysorbate 20 (0.5 ml/liter of Wash Buffer).

NOTE: Polysorbate 20 is a viscous liquid and cannot be measured by a regular pipette. A positive displacement pipette or a syringe should be used to deliver small quantities accurately.

Sample Preparation

Heterogeneous mixtures such as lavage fluids and aspirates may contain contaminants which can interfere in the assay. It is best to check for interference before embarking on a large number of sample measurements. To test for interference, dilute one or two test samples to at least two different dilutions between approximately 20 and 500 pg/ml (*i.e.*, between 20-80% B/B_0). If the two different dilutions of the sample show good correlation (differ by 20% or less) in the final calculated CysLT concentration, purification is not required.

General Precautions

- All samples must be free of organic solvents prior to assay.
- Samples should be assayed immediately after collection; samples that cannot be assayed immediately should be stored at -80°C. Samples containing BSA or FBS may interfere with this assay. AEBSF (Pefabloc SC®) and PMSF inhibit acetylcholinesterase. Samples containing these protease inhibitors should not be used in this assay.
- Samples of mouse and rat origin may contain antibodies which interfere with the assay by binding to goat anti-mouse plate. We recommend that all mouse and rat samples be purified prior to use in the assay.

Lavage Fluids and Aspirates

CysLTs can accumulate to relatively high concentrations in the effusion fluids associated with inflammation (*e.g.*, ascites fluid, synovial fluid, pleural effusion, pericardial or cerebral intraventricular aspirates). Since leukotriene metabolism is incomplete in these circumstances, substantial amounts of LTC_4, LTD_4, and LTE_4 may be present (*e.g.*, bronchoalveolar lavage fluid from asthmatic subjects may contain 700-1,000 pg/ml CysLTs comprised mainly of LTC_4 and LTD_4).[7] Consequently, analysis of these fluids is the optimal application of this assay. Due to the complex nature of these samples there may be significant interference in unpurified samples.[7,8] We have found that we could obtain a good correlation (differ by <20%) in at least two dilutions in the final calculated CysLT concentration in OVA-challenged mouse lavage fluid that was diluted to at least three different dilutions starting at 1:4.

Urine

CysLTs are excreted in urine as intact LTE_4 (~9-12%) and LTE_4 metabolites. Since LTC_4 and LTD_4 are virtually absent from urine, CysLT measurements in urine samples can be accomplished by measuring LTE_4 specifically (LTE_4 ELISA Kit, Item No. 501060). The levels of LTE_4 reported in urine vary depending on the method of measurement. Values obtained from HPLC (50-80 pg/mg creatinine)[7], MS (25-40 pg/mg creatinine),[9] or HPLC followed by ELISA (20-160 pg/mg creatinine)[10] are generally lower than values from ELISA measurements alone (200-1,500 pg/mg creatinine).[11,12]

Culture Medium

Cultured cells synthesizing LTC_4 will generally release it into the medium where it will accumulate without further metabolism. Thus, samples of this type are best analyzed by the measurement of LTC_4 specifically (LTC_4 ELISA Kit, Item No. 501070). Culture medium, in general, can be assayed following dilution with Immunoassay Buffer A. For best results, the standard curve should also be diluted in culture medium.

Preparation of Assay-Specific Reagents

Cysteinyl Leukotriene Express ELISA Standard

Equilibrate a pipette tip in ethanol by repeatedly filling and expelling the tip with ethanol several times. Using the equilibrated pipette tip, transfer 100 µl of the CysLT Express ELISA Standard (Item No. 420504) into a clean test tube, then dilute with 900 µl UltraPure water. The concentration of this solution (the bulk standard) will be 10 ng/ml.

NOTE: If assaying culture medium samples that have not been diluted with at least 10-fold Immunossay Buffer A, culture medium should be used in place of Immunossay Buffer A for dilution of the standard curve.

To prepare the standard for use in ELISA: Obtain eight clean test tubes and number them #1 through #8. Aliquot 900 µl Immunoassay Buffer A to tube #1 and 500 µl Immunoassay Buffer A to tubes #2-8. Transfer 100 µl of the bulk standard (10 ng/ml) to tube #1 and mix thoroughly. Serially dilute the standard by removing 500 µl from tube #1 and placing in tube #2; mix thoroughly. Next, remove 500 µl from tube #2 and place it into tube #3; mix thoroughly. Repeat this process for tubes #4-8. These diluted standards should not be stored for more than 24 hours.

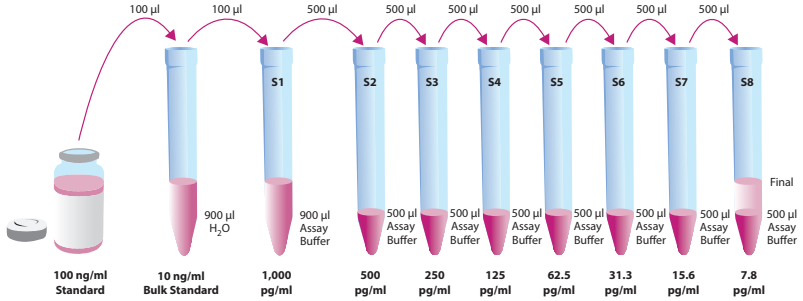


Figure 5. Preparation of the CysLT standards

Cysteinyl Leukotriene Express AChE Tracer

Reconstitute the CysLT Express AChE Tracer as follows:

100 dtn CysLT Express AChE Tracer (96-well kit; Item No. 10011323): Reconstitute with 6 ml Immunoassay Buffer A.

OR

500 dtn CysLT Express AChE Tracer (480-well kit; Item No. 10011323): Reconstitute with 30 ml Immunoassay Buffer A.

Transfer the reconstituted tracer to a polypropylene tube or vial, store at 4°C (*do not freeze*), and use within one week. A 20% surplus per vial of tracer has been included to account for any incidental losses.

NOTE: The 480-well size kit contains two 500 dtn vials of tracer so that the kit may be used for more than one week. Each vial of tracer will be stable for one week from reconstitution.

Cysteinyl Leukotriene Express ELISA Monoclonal Antibody

Reconstitute the CysLT Express ELISA Monoclonal Antibody as follows:

100 dtn CysLT Express ELISA Monoclonal Antibody (96-well kit; Item No. 10011322): Reconstitute with 6 ml Immunoassay Buffer A.

OR

500 dtn CysLT Express ELISA Monoclonal Antibody (480-well kit; Item No. 10011322): Reconstitute with 30 ml Immunoassay Buffer A.

Store the reconstituted CysLT Express ELISA Monoclonal Antibody at 4°C. It will be stable for at least four weeks. A 20% surplus of antibody has been included to account for any incidental losses.

Plate Set Up

The 96-well plate(s) included with this kit is supplied ready to use. It is not necessary to rinse the plate(s) prior to adding the reagents. *NOTE: If you do not need to use all the strips at once, place the unused strips back in the plate packet and store at 4°C. Be sure the packet is sealed with the desiccant inside.*

Each plate or set of strips must contain a minimum of two blanks (Blk), two non-specific binding wells (NSB), two maximum binding wells (B_0), and an eight point standard curve run in duplicate. *NOTE: Each assay must contain this minimum configuration in order to ensure accurate and reproducible results.* Each sample should be assayed at two dilutions and each dilution should be assayed in duplicate. For statistical purposes, we recommend assaying samples in triplicate.

A suggested plate format is shown in Figure 6, below. The user may vary the location and type of wells present as necessary for each particular experiment. The plate format provided below has been designed to allow for easy data analysis using a convenient spreadsheet offered by Cayman (see page 22, for more details). We suggest you record the contents of each well on the template sheet provided (see page 30).

	1	2	3	4	5	6	7	8	9	10	11	12
A	Blk	S1	S1	1	1	1	9	9	9	17	17	17
B	Blk	S2	S2	2	2	2	10	10	10	18	18	18
C	NSB	S3	S3	3	3	3	11	11	11	19	19	19
D	NSB	S4	S4	4	4	4	12	12	12	20	20	20
E	B_0	S5	S5	5	5	5	13	13	13	21	21	21
F	B_0	S6	S6	6	6	6	14	14	14	22	22	22
G	B_0	S7	S7	7	7	7	15	15	15	23	23	23
H	TA	S8	S8	8	8	8	16	16	16	24	24	24

Blk - Blank
TA - Total Activity
NSB - Non-Specific Binding
B_0 - Maximum Binding
S1-S8 - Standards 1-8
1-24 - Samples

Figure 6. Sample plate format

Performing the Assay

Pipetting Hints

- Use different tips to pipette each reagent.
- Before pipetting each reagent, equilibrate the pipette tip in that reagent (*i.e.*, slowly fill the tip and gently expel the contents, repeat several times).
- Do not expose the pipette tip to the reagent(s) already in the well.

Addition of the Reagents

1. **Immunoassay Buffer A**

   Add 100 µl Immunoassay Buffer A to NSB wells. Add 50 µl Immunoassay Buffer A to B_0 wells. If culture medium was used to dilute the standard curve, substitute 50 µl of culture medium for Immunoassay Buffer A in the NSB and B_0 wells (*i.e.*, add 50 µl culture medium to NSB and B_0 wells and 50 µl Immunoassay Buffer A to NSB wells).

2. **CysLT Express ELISA Standard**

   Add 50 µl from tube #8 to both of the lowest standard wells (S8). Add 50 µl from tube #7 to each of the next two standard wells (S7). Continue with this procedure until all the standards are aliquoted. The same pipette tip should be used to aliquot all the standards. Before pipetting each standard, be sure to equilibrate the pipette tip in that standard.

3. **Samples**

   Add 50 µl of sample per well. Each sample should be assayed at a minimum of two dilutions. Each dilution should be assayed in duplicate (triplicate recommended).

4. **CysLT Express AChE Tracer**

   Add 50 µl to each well *except* the TA and the Blk wells.

5. **CysLT Express ELISA Monoclonal Antibody**

   Add 50 µl to each well *except* the TA, the NSB, and the Blk wells.

Well	Immunoassay Buffer A	Standard/Sample	Tracer	Antibody
Blk	-	-	-	-
TA	-	-	5 µl (at devl. step)	-
NSB	100 µl	-	50 µl	-
B_0	50 µl	-	50 µl	50 µl
Std/Sample	-	50 µl	50 µl	50 µl

Table 1. Pipetting summary

Incubation of the Plate

Cover each plate with plastic film (Item No. 400012) and incubate for two hours at room temperature on an orbital shaker.

Development of the Plate

1. Reconstitute Ellman's Reagent immediately before use (20 ml of reagent is sufficient to develop 100 wells):

   100 dtn vial Ellman's Reagent (96-well kit; Item No. 400050): Reconstitute with 20 ml of UltraPure water.

   OR

   250 dtn vial Ellman's Reagent (480-well kit; Item No. 400050): Reconstitute with 50 ml of UltraPure water.

NOTE: Reconstituted Ellman's Reagent is unstable and should be used the same day it is prepared; protect the Ellman's Reagent from light when not in use. Extra vials of the reagent have been provided should a plate need to be re-developed or multiple assays be run on different days.

2. Empty the wells and rinse five times with Wash Buffer.
3. Add 200 µl of Ellman's Reagent to each well
4. Add 5 µl of tracer to the TA wells.
5. Cover the plate with plastic film. Optimum development is obtained by using an orbital shaker equipped with a large, flat cover to allow the plate(s) to develop in the dark. This assay typically develops (*i.e.*, B_0 wells ≥0.3 A.U. (blank subtracted)) in 60-90 minutes.

Reading the Plate

1. Wipe the bottom of the plate with a clean tissue to remove fingerprints, dirt, etc.
2. Remove the plate cover being careful to keep Ellman's Reagent from splashing on the cover. *NOTE: Any loss of Ellman's Reagent will affect the absorbance readings. If Ellman's Reagent is present on the cover, use a pipette to transfer the Ellman's Reagent into the well. If too much Ellman's Reagent has splashed on the cover to easily redistribute back into the wells, wash the plate three times with wash buffer and repeat the development with fresh Ellman's Reagent.*
3. Read the plate at a wavelength between 405 and 420 nm. The absorbance may be checked periodically until the B_0 wells have reached a minimum of 0.3 A.U. (blank subtracted). The plate should be read when the absorbance of the B_0 wells in the range of 0.3-1.5 A.U. (blank subtracted). If the absorbance of the wells exceeds 2.0, wash the plate, add fresh Ellman's Reagent and let it develop again.

ANALYSIS

Many plate readers come with data reduction software that plots data automatically. Alternatively a spreadsheet program can be used. The data should be plotted as either %B/B_0 *versus* log concentration using a four-parameter logistic fit or as logit B/B_0 *versus* log concentration using a linear fit. *NOTE: Cayman has a computer spreadsheet available for data analysis. Please contact Technical Service or visit our website (www.caymanchem.com/analysis/elisa) to obtain a free copy of this convenient data analysis tool.*

Calculations

Preparation of the Data

The following procedure is recommended for preparation of the data prior to graphical analysis.

NOTE: If the plate reader has not subtracted the absorbance readings of the blank wells from the absorbance readings of the rest of the plate, be sure to do that now.

1. Average the absorbance readings from the NSB wells.
2. Average the absorbance readings from the B_0 wells.
3. Subtract the NSB average from the B_0 average. This is the corrected B_0 or corrected maximum binding.
4. Calculate the B/B_0 (Sample or Standard Bound/Maximum Bound) for the remaining wells. To do this, subtract the average NSB absorbance from the S1 absorbance and divide by the corrected B_0 (from Step 3). Repeat for S2-S8 and all sample wells. (To obtain %B/B_0 for a logistic four-parameter fit, multiply these values by 100.)

NOTE: The TA values are not used in the standard curve calculations. Rather, they are used as a diagnostic tool; the corrected B_0 divided by the actual TA (10X measured absorbance) will give the %Bound. This value should closely approximate the %Bound that can be calculated from the ***Sample Data*** *(see page 24). Erratic absorbance values and a low (or no) %Bound could indicate the presence of organic solvents in the buffer or other technical problems (see page 28 for* ***Troubleshooting****).*

Plot the Standard Curve

Plot %B/B_0 for standards S1-S8 *versus* CysLT concentration using linear (y) and log (x) axes and perform a 4-parameter logistic fit.

Alternative Plot - The data can also be lineraized using a logit transformation. The equation for this conversion is shown below. *NOTE: Do not use %B/B_0 in this calculation.*

$$\text{logit } (B/B_0) = \ln [B/B_0/(1 - B/B_0)]$$

Plot the data as logit (B/B_0) *versus* log concentrations and perform a linear regression fit.

Determine the Sample Concentration

Calculate the B/B_0 (or %B/B_0) value for each sample. Determine the concentration of each sample by identifying the %B/B_0 on the standard curve and reading the corresponding values on the x-axis. *NOTE: Remember to account for any concentration of the sample prior to the addition to the well.* Samples with %B/B_0 values greater than 80% or less than 20% should be re-assayed as they generally fall out of the linear range of the standard curve.

Performance Characteristics

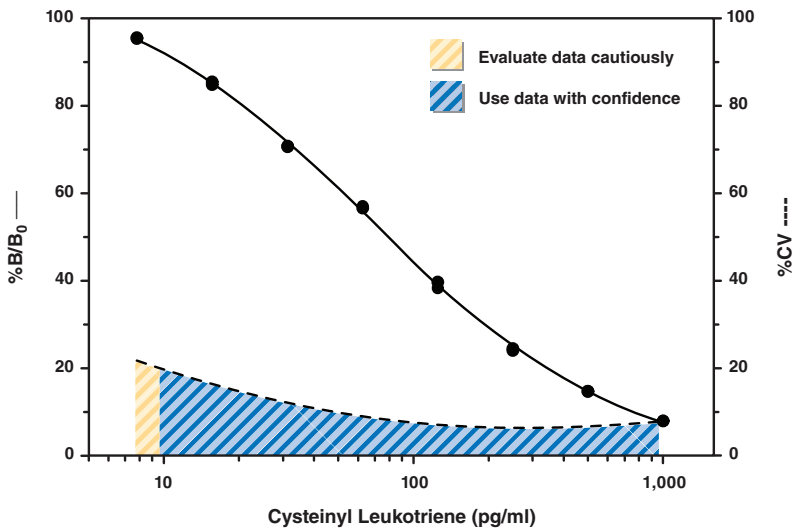
Sample Data

The standard curve presented here is an example of the data typically produced with this kit; however, your results will not be identical to these. You must run a new standard curve. Do not use the data below to determine the values of your samples. Your results could differ substantially.

	Raw Data		Average	Corrected
Total Activity	1.450	1.498	1.454	
NSB	0.013	0.003	0.008	
B_0	0.754	0.759		
	0.782	0.823	0.780	0.772

Dose (pg/ml)	Raw Data		Corrected		%B/B_0	
1,000	0.069	0.069	0.061	0.061	7.8	7.8
500	0.120	0.120	0.112	0.112	14.5	14.5
250	0.193	0.196	0.185	0.188	23.9	24.3
125	0.313	0.302	0.305	0.294	39.5	38.1
62.5	0.446	0.444	0.438	0.436	56.8	56.4
31.3	0.551	0.552	0.543	0.544	70.4	70.5
15.6	0.666	0.661	0.658	0.653	85.2	84.6
7.8	0.743	0.744	0.735	0.736	95.1	95.3

Table 2. Typical results



Assay Range = 7.8-1,000 pg/ml
Sensitivity (defined as 80% B/B_0) = 20 pg/ml
Mid-point (defined as 50% B/B_0) = 70-110 pg/ml

The sensitivity and mid-point were derived from the standard curve shown above. Cysteinyl Leukotriene was diluted with Immunoassay Buffer A.

Figure 6. Typical standard curve

Precision:

The intra- and inter-assay CVs have been determined at multiple points on the standard curve using Immunoassay Buffer A as the sample matrix. These data are summarized in the graph on page 25 and in the table below.

Dose (pg/ml)	%CV* Intra-assay variation	%CV* Inter-assay variation
1,000	5.6	16.5
500	7.1	9.8
250	9.2	10.3
125	8.8	6.1
62.5	6.6	9.3
31.3	11	11.3
15.6	12.4	19.1
7.8	24	†

Table 3. Intra- and inter-assay variation

*%CV represents the variation in concentration (not absorbance) as determined using a reference standard curve.

†Outside of the recommended usable range of the assay.

Cross Reactivity:

Compound	Cross Reactivity
N-methyl Leukotriene C_4	124%
Leukotriene C_4	100%
Leukotriene D_4	100%
Leukotriene E_4	65%
Arachidonic Acid	<0.01%
5,6-DiHETE	<0.01%
5,15-DiHETE	<0.01%
5-HETE	<0.01%
12-HETE	<0.01%
15-HETE	<0.01%
Leukotriene B_4	<0.01%
Prostaglandin D_2	<0.01%
Prostaglandin E_2	<0.01%

Table 4. Cross Reactivity of the CysLT Express ELISA

RESOURCES

Troubleshooting

Problem	Possible Causes	Recommended Solutions
Erratic values; dispersion of duplicates	A. Trace organic contaminants in the water source B. Poor pipetting/technique	Replace activated carbon filter or change source of UltraPure Water
High NSB (>10% of B_0)	A. Poor washing B. Exposure of NSB wells to specific antibody	Rewash plate and redevelop
Very low B_0	A. Trace organic contaminants in the water source B. Plate requires additional development time C. Dilution error in preparing reagents	A. Replace activated carbon filter or change source of UltraPure Water B. Return plate to shaker and re-read later
Low sensitivity (shift in dose response curve)	Standard is degraded	Replace standard
Analyses of two dilutions of a biological sample do not agree (*i.e.*, more than 20% difference)	Interfering substances are present	Samples may not be suitable for use in this assay
Only Total Activity (TA) wells develop	Trace organic contaminants in the water source	Replace activated carbon filter or change source of UltraPure Water

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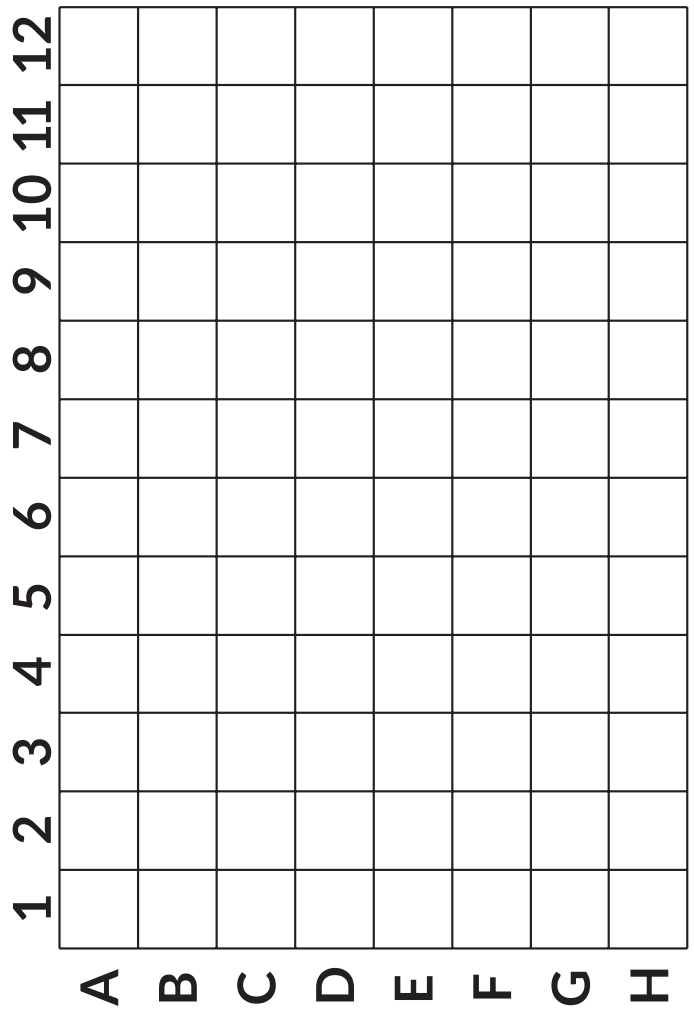

NOTES

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